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# Exercise and cardioprotection against ischemia reperfusion injury: A Review

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Cardiovascular disease (CVD) has been the leading cause of death in the United States for nearly a century. Despite relatively recent progress in cardiovascular health interventions, CVD and particularly ischemic heart disease remain a significant global health problem. Various approaches to evoke a preconditioned phenotype has largely been unsuccessful, highlighting several logistical problems in pharmacologic approaches to cardioprotection. Issues include the unpredictable nature of ischemic events, the cost of chronically delivering pharmacologic countermeasures, and the fact that many exogenous stimuli to cardiac preconditioning result in rapid cellular habituation and a waning of cardioprotection. Accordingly, many lifestyle approaches to cardiac preconditioning have been investigated, including moderate intensity aerobic exercise. Regular exercise represents a pragmatic, sustainable, and cost-effective measure for reducing CVD risk and for provoking robust levels of cardioprotection against ischemic events. The scientific sub-field borne from this research line is known as exercise-induced cardiac preconditioning. Key discoveries of the nature of exercise preconditioning reveal that clinically relevant protection can be obtained with moderate intensity exercise in as little as 1-3 days and occurs independent of sex and age. The short-term nature of the exercise preconditioning stimulus highlights the importance of biochemical alterations as a key mechanistic process for this protection. Importantly, mechanisms central to exercise-induced cardiac preconditioning include a concert of up-regulated, or allosterically altered, factors that mitigate calcium overload, free radical damage and oxidative stress, and bioenergetic dyshomeostasis. Finally, the efficacy of exercise-induced cardiac preconditioning highlights the value of repetitive physical activity as a scientific approach to drug discovery.

Keywords: exercise, cardioprotection; ischemia-reperfusion injury; preconditioning; ischemic heart disease

#### 1. Introduction

Cardiovascular disease (CVD), including ischemic heart disease, remains a leading form of morbidity and mortality across the world (Virani et al., 2020). In response, scientists continue to search for counter measures to CVD that are simultaneously pragmatic, cost effective, and sustainable. One such approach includes the decades-long attempt to reverse engineer preclinical understanding of biochemical cardioprotection into a pharmacological counter therapy against CVD insults. Efforts to date, however, have been thwarted by the faceted and evolutionary nature of CVD, the inability to prognosticate ischemic events, and a Quixotic search for an exogenous pharmacologic agent that will produce lasting cardioprotection without cellular habituation (Bolli and Marban, 1999; Quindry and Franklin, 2018). In the absence of a viable pharmacologic solution to counter CVD incidence and severity, physicians and scientists have also explored lifestyle interventions, including a cardiovascular exercise regimen. Indeed, exercise and other healthy lifestyle counter therapies precondition the myocardium

against a host of pathological stressors, including ischemia.

The cardioprotective nature of aerobic exercise is wellestablished (Paffenbarger et al., 1966; Powers et al., 1998; Starnes et al., 2003). Moreover, while structured exercise (e.g. 3-5 days/week, 30-60 minutes in duration, at an intensity of 50%-85% heart rate reserve) clearly improves cardiovascular health and preconditions the heart against CVD (Garber et al., 2011), accumulated amounts of less regimented physical activity are also protective (Riebe et al., 2018). Initial evidence in this regard was observed more than half a century ago from epidemiological studies, which demonstrated a relationship between occupational physical activity and cardiovascular disease outcomes (Paffenbarger et al., 1966). Relevant to the current discussion, this early epidemiological study of cardioprotection provided an almost prescient conclusion (Paffenbarger et al., 1966):

"The association between work activity and coronary mortality, when considered with the lack of such association with stroke mortality, suggests that physical

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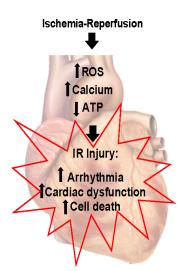


Figure 1. Ischemia reperfusion injury. Ischemia-reperfusion results in arrhythmia, decreased cardiac function, and cell death. Elevated free radical production, calcium overload, and decreased mitochondrial function are central to ischemia-reperfusion injury. Figure created by modification of an image created by Patrick J. Lynch, medical illustrator; C. Carl Jaffe, MD, cardiologist. https://creativecommons. org/licenses/by/2.5/

## activity influences the myocardium or its function more than the atherosclerotic process."

Indeed, the capacity of repetitive muscular work (formal exercise or plentiful physical activity) to induce biochemical cardiac preconditioning was so powerful that it was apparent to epidemiologists of the mid 20<sup>th</sup> century.

Over the past few decades, much has been uncovered about the mechanisms of this protection. In addition to decreasing risk factors associated with the development of cardiovascular diseases, regular exercise directly protects against cardiac ischemia- reperfusion (IR) injury via cellular biochemical alterations (Demirel et al., 2001; Lennon et al., 2004a; Quindry et al., 2005; Quindry et al., 2012). Thus, given the fact that exercise can precondition the myocardium indefinitely, recent scientific interest has been directed at uncovering mechanisms of exercise cardioprotection for the purpose of designing pharmaceuticals aimed at mitigating damage from IR injury. As is the case in many applied lines of biomedical research, much of the fundamental knowledge about exercise induced cardioprotection against CVD is derived from animal studies, primarily using rodent models. Moreover, while exercise improves outcomes in models of heart failure, vascular dysfunction, and metabolic disease, the largest body of work relates to ischemic heart disease (Powers et al., 1998; Demirel et al., 2001; Starnes et al., 2003; Lennon et al., 2004a; Quindry et al., 2005; Quindry et al., 2012). Accordingly, the thrust of this review is directed toward the latter, primarily detailing what is known about exercise preconditioning against myocardial infarction. Moreover, the topic of exercise induced cardioprotection has been reviewed elsewhere with an emphasis toward scientific understanding of signaling pathways (Starnes and Taylor, 2007; Penna et al., 2020), as a unique scientific approach (Brown and Moore, 2007; Powers et al., 2014; Quindry, 2017; Wojcik et al., 2018), and as an integrated pharmacologic and rehabilitative approach to the treatment and prevention of cardiovascular disease (Fiuza-Luces et al., 2018; Quindry and Franklin, 2018; Thijssen et al., 2018; Chowdhury et al., 2019). The current review updates these perspectives within the more holistic narrative of exercise as a potent means of conditioning medicine for improved heart health in those at significant risk for an ischemic event.

## Experimental paradigms of exercise preconditioning against myocardial ischemia

Animal models used to investigate exercise induced cardioprotection, commonly performed in rats and mice, include various exercise protocols followed by surgically induced IR *in vivo*, or global ischemia applied *ex vivo*. Although protocols have varied in both the exercise volume employed and duration of IR investigated, overall, many studies have employed exercise volumes comparable to human exercise capacity and measurement of clinically relevant outcomes observed in humans. Table 1 provides a description of a common study design for experimental models of exercise induced cardioprotection. The reader is further directed to a recent 'systematic comparison of exercise training protocols on animal models of cardiovascular capacity' for a more in depth presentation of exercise protocols (Feng et al., 2019).

Scientific efficacy of these preclinical studies, of course, is reliant upon the applicability of the exercise interventions in addition to the clinical relevance of the infarct challenge and key dependent outcome measures of cardiac pathology. To this end, it is essential to recognize that the exercise prescriptions used for improving cardiovascular health in humans are proportionally scaled to the animal model of interest. Specifically, the American College of Sports Medicine 10<sup>th</sup> edition of Guidelines for Exercise Testing and Prescription can be redefined for exercise frequency, intensity, duration, modality, and progression in animal studies (Riebe et al., 2018). Indeed, the exercise frequency of 3-5 days/week of aerobic exercise is common to both human exercise prescriptions and rodent exercise models (Criswell et al., 1993; Powers et al., 1998; McGinnis et al., 2015; Riebe et al., 2018). Accordingly, while the modes of continuous aerobic exercise involve forced treadmill exercise (typically with an electrified shock grid on the tail end of the running surface) or immersive swimming, the modalities do parallel human physical activity. Similarly, the metabolic costs of these activities can be accurately quantified in both humans and animals using applications of indirect calorimetry (Lawler et al., 1993; Riebe et al., 2018). Based on this understanding, the exercise intensity prescribed in exerciseinduced cardioprotection studies spans the identical intensity window used in humans (Bowles et al., 1992; Starnes et al., 2003; Lennon et al., 2004a; Riebe et al., 2018).

It is worth acknowledging that forced exercise in rodent models is a potentially confounding aspect of most exerciseinduced cardioprotection studies (Mancardi et al., 2009). Fortunately, however, evidence from investigations of rodentselected wheel running support the application of human exercise prescriptions in animals, noting that when mice are provided the chance to exercise at their volition on a running wheel, they will naturally accumulate a volume of running that equals the prescribed distance outlined in experimental models of exercise (Lightfoot et al., 2010). In aggregate, these findings provide validation to the common exercise approaches used to evoke a cardioprotective phenotype against ischemic insults.

As mentioned previously, clinically relevant outcomes and key dependent measures are essential to understanding the role of exercise and an ischemic-resistant heart. An overview of the pathology of IR has been summarized in Figure 1. Given that myocardial ischemia results in an evolutionary pathology, common clinical endpoints are needed. Clinically relevant outcomes can be most comprehensively examined in surgical ligation models *in vivo*. In these studies, electrocardiogram (ECG) analysis is readily quantified during both ischemia and reperfusion, and ventricular ectopy can be tracked for potentially lethal arrhythmias using clinical scoring systems based on Lambeth convention dictates (Curtis and Walker, 1988; Miller et al., 2012). Similarly, ventricular pump function can be precisely quantified by pulse pressure measurements Table 1. Methodological approach common to exercise-induced cardioprotection research studies using rodents and other mammals.

Animals randomized to groups	Day 1-5: Treadmill Habituation	Day 6-7: Rest	Day 8 - 10: Treatment	Day 11: Surgical IR	Exercise preconditioning outcomes
Exercise group	Begin with 10 min treadmill running, increase 10min per day for 5 days	No treadmill exposure	30-60 min treadmill running 50 - 75% VO2max 3 consecutive days	50 min Ischemia, 120 min Reperfusion with functional analysis, followed by post- mortem histochemical and biochemical analysis	↑ cardiac function ↓ infarct area ↓ serum damage markers ↓ tissue-level damage
Sedentary group	Begin with 10 min motionless treadmill, increase 10 min per day for 5 days	No treadmill exposure	30-60 min motionless treadmill 3 consecutive days		

from indwelling arterial pressure catheters, pressure volume loop analysis, transthoracic echocardiography, and isolated perfused working heart preparations (Hamilton et al., 2001; Quindry et al., 2005; Starnes et al., 2005). Perhaps most commonly examined, infarct size can be accurately quantified by examining the release of cardiac-specific enzymes from blood plasma or cardiac perfusate collected from isolated heart preparations (Hamilton et al., 2003; Lennon et al., 2004a). In addition, post-mortem histological cross-sections can be used to track both the ischemic area and the infarct region produced by surgical ligation of the left anterior descending artery (LAD) (Hamilton et al., 2003; Quindry et al., 2007; Quindry et al., 2012). Finally, post-mortem examination of the ventricular muscle can be examined for biochemical alterations such that the ischemic area can be compared to perfused regions of the same heart. This simple, but powerful, scientific approach provides invaluable insights to the biochemical alterations in hearts from exercised and sedentary treatments (Quindry et al., 2010a; McGinnis et al., 2015; Miller et al., 2015). As will be discussed subsequently, this approach to using each heart as its own control has been foundational in the early studies to suggest that exercise training appears to cardioprotect through subtle modifications in cellular processes such as apoptosis and autophagy (Quindry et al., 2005; Quindry et al., 2007; Quindry et al., 2012; McGinnis et al., 2015; Miller et al., 2015).

#### Efficacy in various populations and required exercise dose

Descriptive evidence supporting the notion of exercise-induced cardiac preconditioning has accumulated over the last 25-30 years. Foundational to this line of work is the question, *is exercise induced cardiac preconditioning effective for both sexes and across the lifespan*? Numerous studies confirm that both males and females exhibit equally robust protection against infarction (Brown et al., 2005a; Chicco et al., 2007). Perhaps most important, given that CVD accrues as a function of advancing age, is evidence from rodent models that exercise induced cardioprotection is effective against ischemic insults in both young and old animals (Starnes et al., 2003; Quindry et al., 2005). Thus, these early studies confirm that exercise induced cardioprotection can be conferred in essentially all hearts, apparently without exclusion based on age or sex.

When it comes to prescribing exercise to prevent or treat CVD, the frequency, type, dose, and intensity of exercise are all important. As mentioned earlier, the methodological approach to exercise training in rodents (and larger mammals) is proportional to exercise prescriptions used to produce improved health and fitness in humans. Accordingly, one would be inclined to ask, *what is the dose of exercise needed to produce a cardioprotected phenotype*?

In regard to the dose of exercise needed to produce a

cardioprotected phenotype, long-term, regular exercise clearly promotes an improved risk factor profile and beneficial alterations in vascularization and structural improvements in ventricular architecture (Gielen et al., 2015). Likewise, regular exercise performed over the course of many weeks results in improved cardiac function and decreased damage and infarct area following IR (Bowles et al., 1992; Powers et al., 1998; Brown et al., 2005b). Interestingly, subsequent investigations demonstrate protection is achieved in as little as 1-3 days of exercise (Demirel et al., 2001; Brown et al., 2005a). That is, exercise preconditioning against ventricular dysrhythmias, decrements in ventricular pump function, and infarct damage occur with just 1-3 daily bouts of exercise prior to IR (Yamashita et al., 1999; Starnes et al., 2003; Taylor et al., 2007; Quindry et al., 2010a; Quindry et al., 2012). Indeed, there is little to no conflicting evidence that short-term bouts (days) of exercise training are sufficient to produce a robustly protected phenotype. Moreover, while the preconditioned state may be achieved within a few days, it important to point out that the protected phenotype is by all accounts sustained with continued participation in exercise (Powers et al., 1998). This finding, while perhaps obvious to some readers, starkly contrasts nonexercise approaches to cardiac preconditioning where cellular habituation of the underlying mechanisms occurs within days (Lefer and Bolli, 2011; Peart et al., 2011).

Quite interestingly, both moderate and high intensity exercise are protective, with the duration of many studies set at 30-60 min per day (Starnes et al., 2003; Lennon et al., 2004a; Quindry et al., 2012; Miller et al., 2015). As discussed earlier, many studies have used exercise interventions that mirror the current baseline physical activity recommendations for adults according to the American College of Sports Medicine, which is moderate intensity exercise for 30-60 min per day 3-5 days per week (Garber et al., 2011; Riebe et al., 2018). This finding is important because it suggests that the 'dose' of exercise is relatively low compared to maximal capacity in humans and that the level of exercise required for cardioprotective benefit is obtainable in various populations/fitness levels. It appears that moderate and high intensity exercise are equally protective (Lennon et al., 2004a) though some evidence indicates there may be a slightly greater level of protection regarding post-ischemic cardiac function with higher intensity exercise (Bowles et al., 1992). Follow up studies are needed to further characterize the effect of intensity on cardioprotective outcomes. Additionally, although most studies have employed moderate intensity aerobic exercise, it appears resistance exercise may also confer cardioprotective benefit (Soufi et al., 2011), although more evidence is needed due to the limited number of studies on this topic. Additionally, the protective nature of acute exercise appears to be threshold dependent,

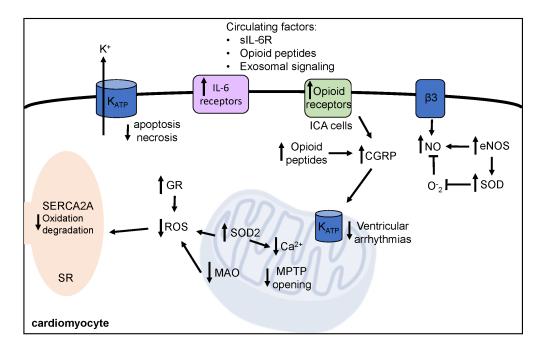


Figure 2. Mechanisms of exercise induced cardioprotection against ischemia-reperfusion injury. Exercise prior to ischemia-reperfusion decreases arrhythmia, improves cardiac function, and decreases cell death. Short-term exercise induces protective biochemical alterations within the cardiomyocyte, which are summarized here. SOD2, mitochondrial isoform of superoxide dismutase; GR, glutathione reductase; MAO, monoamine oxidase; MPTP, mitochondrial permeability transition pore; NO, nitric oxide; eNOS, endothelial nitric oxide synthase; SERCA2A, sarcoplasmic endoplasmic calcium ATPase; K<sub>ATP</sub>, ATP-sensitive potassium channels; IL-6, interleukin-6, sIL-6R, soluble IL-6 receptor; ICA, intrinsic cardiac adrenergic; CGRP, reticulum calcitonin gene related peptide.

where longer exercise bouts do not necessarily translate to greater protection (Lennon et al., 2004a). The relatively short time-course needed to obtain clinically relevant protection underscores the notion that the primary mechanisms of exercise cardiac preconditioning are biochemical in nature and can occur independent of cardiac remodeling.

## Limitations to animal models of exercise induced cardiac preconditioning

Perhaps needless to state, animal models of exercise induced cardiac preconditioning allow for invasive measurements that would be difficult and/or impossible in human populations. Despite the clear benefits animal models provide, it is important to note significant inherent limitations. Many studies use rodents that are free from CVD prior to surgically induced IR, which makes direct comparisons to the human population where CVD is present difficult. Although support exists for the presence of exercise induced cardiac preconditioning in humans, as will be discussed in a later section, it must be acknowledged that the healthy heart may respond differently to an exercise stress as compared to an unhealthy heart. However, some studies have investigated exercise induced cardiac preconditioning in disease models. For example, exercise is cardioprotective against the progression of heart failure in an animal model of left ventricular hypertrophy (Marshall et al., 2013). Additionally, exercise remains effective at conferring protection in the presence of traditional risk factors for CVD such as obesity, elevated blood glucose, insulin, and cholesterol levels (Pons et al., 2013; Lund et al., 2015). Finally, co-morbidity disease models, combined with a surgically induced ischemic challenge, confirm the potent role of exercise to cardioprotect despite the presence of uncontrolled diabetes (Kupai et al., 2015).

Although exercise appears to be equally cardioprotective in male and females, mechanistic differences may exist (Brown et al., 2005a; Chicco et al., 2007). While many studies have used female animals or both male and female animals, a large portion of studies have used only male animals, therefore it

is reasonable to hypothesize that undiscovered mechanistic differences between sexes likely remain. Lastly, as CVD and ischemic heart disease increase with advancing age, investigations including aged animal models are necessary. Although evidence suggests exercise induced cardiac preconditioning remains effective in older animals (Starnes et al., 2003; Quindry et al., 2005), many studies have not used older animals. Moreover, it cannot be overlooked that in the 2005 investigation by Quindry and colleagues, where TUNEL staining was used as a surrogate of apoptotic processes in the heart, the level of TUNEL-positive nuclei in unstressed aged animals was elevated in an age-dependent fashion. While the exercise intervention was short-term (days), there is no reason to believe that long-term exercise training would have completely mitigated the age-dependent apoptotic process in the heart (Quindry et al., 2005). In total, it is encouraging to note that exercise acutely preconditions the aged heart, but that in the context of conditioning medicine, where most individuals are not life-long exercise habituated, the intervention is not likely to reverse decades of aging and disuse. Indeed, this fact underpins the modern call to action to investigate exercise preconditioning in aged animal models.

## Mechanisms of exercise induced cardioprotection against an ischemic insult

It is well established that acute, (1-3 days) moderate intensity exercise improves outcomes following IR. Specifically, exercised hearts maintain greater function by preserved contractility (Starnes et al., 2003; Lennon et al., 2004a; Quindry et al., 2005), better maintenance of systolic blood pressure (Demirel et al., 2001; Hamilton et al., 2003), and decreased occurrence of arrhythmias (Quindry et al., 2010a; Miller et al., 2012; McGinnis et al., 2015). Exercise preconditioning also results in smaller infarct area, decreased levels of apoptotic cell death, and circulating markers of cardiac tissue damage (Lennon et al., 2004a; Quindry et al., 2007; McGinnis et al., 2015; Miller et al., 2015). Given the short-term nature of the robust protection afforded by exercise preconditioning, biochemical mechanisms that quickly respond to the exercise stimulus are of particular interest and include alterations in antioxidant capacity, ion channel function, and various signaling molecules and pathways. Current understanding of these mechanisms is reviewed in the following sections with a summary provided in Figure 2.

#### Antioxidants

Increased free radical production and elevated intracellular calcium levels occur during myocardial IR, resulting in cellular dysfunction and damage (Zweier, 1988; Grill et al., 1992). Limiting the production of free radicals and resulting oxidative stress protects against cell death and dysfunction during IR (Grill et al., 1992). Exercise is well documented to result in upregulated antioxidant defenses, which represents a primary mechanism for limiting cell dysfunction and death resulting from IR (French et al., 2008). Superoxide dismutases (SOD) provide the primary cellular defenses against free radical accumulation by conversion of superoxide to hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) (Fridovich, 1997). Various isoforms of SOD have been identified, including copper/zinc SOD, mitochondrial manganese SOD (MnSOD or SOD2), and extracellular SOD (ecSOD) (Fridovich, 1997). Relative to the exercised heart, the mitochondrial isoform of superoxide dismutase (SOD2 or MnSOD) has been consistently shown to be upregulated following cardioprotective exercise, and to be essential for exercise induced cardioprotection (French et al., 2008). Importantly, the essentiality of SOD2 to the exercised phenotype has been clearly demonstrated through the application of antisense oligonucleotides directed against the endogenous antioxidant. Given the importance of SOD2 to long-term muscle function and health, these knockdown approaches from several groups indicate that a relatively modest (20%-50%) increase in SOD2 expression is protective against ventricular dysrhythmias and tissue infarct, but ventricular pump function (Yamashita et al., 1999; Hamilton et al., 2003; Lennon et al., 2004a). Extracellular SOD increases in response to exercise in the aortic endothelium, which is mediated by endothelial nitric oxide synthase (eNOS) (Fukai et al., 2000). The elevated ecSOD preserves nitric oxide (NO), and may contribute to exercise-derived cardioprotection (Fukai et al., 2000), as will be described in the next section. However, the role of extracellular ecSOD is unstudied in this context.

Early studies indicated that exercise training improves calcium handling during IR (Bowles et al., 1992). Although conflicts in the literature exist, some evidence indicates that exercise training improves mitochondrial tolerance to calcium overload, resulting in decreased mitochondrial permeability transition pore (mPTP) opening (Marcil et al., 2006; Starnes et al., 2007; Magalhaes et al., 2014). In addition, increased mitochondrial antioxidant concentration limits calcium overload that occurs during IR (Buja, 2005; Starnes et al., 2007). Indeed, exercise training increases SOD2, protecting against oxidative modification and calpain-mediated degradation of calcium handling proteins such as L-type calcium channels, phospholamban, and sarcoplasmic/endoplasmic reticulum calcium ATPase, which improves cardiac function and decreases tissue necrosis and apoptosis following IR (French et al., 2006; French et al., 2008). While these studies provide convincing evidence of the role of improved calcium handling as a key component of exercise-induced cardiac preconditioning, this area remains understudied relative to exercise-induced cardioprotection against ischemic insults.

Given that endogenous antioxidants are typically held to function as a network defense, conversion of  $H_2O_2$  produced by SOD2 is likely to be just as important as the quenching of  $O_2$ -

directly. To this end, both the endogenous antioxidant catalase and the glutathione system, which function downstream of SOD to convert  $H_2O_2$  to  $H_2O$  and  $O_2$ , have been extensively investigated. Catalase is inconsistently upregulated in exercised hearts, suggesting that components of the glutathione system may be an essential collaborator to SOD2 (Lennon et al., 2004b; Lennon et al., 2004a; French et al., 2008; Dao et al., 2011). Accordingly, myocardial glutathione reductase activity is enhanced following exercise and appears to serve as the essential second step for exercise induced cardioprotection for free radical generation of superoxide from the mitochondria during an IR challenge (Frasier et al., 2011). However, it appears that the other facets of the glutathione system do not need to up-regulate in parallel. Indeed, mixed results exist in terms of upregulation and requirement for protection by glutathione peroxidase, and it is proposed that improved ability to replenish glutathione by glutathione reductase (as opposed to increased glutathione peroxidase content) may be the mechanism responsible for glutathione system's role in exercise cardioprotection (Frasier et al., 2011).

With regard to endogenous antioxidant function, it is important to note that just as IR pathology occurs through the parallel events of free radical production and calcium dyshomeostasis, exercise-induced protection mediates simultaneous protection through redundant mechanisms that offer tandem levels of protection. For instance, upregulation of antioxidant defense systems, and in particular SOD2, appear to benefit myocardial calcium handling following IR. It is currently unknown whether these observations are the result of directed antioxidant events in the cytosol or are due to the upstream quenching of free radical chain reactions. Nonetheless, it is clear that modulation of endogenous antioxidants, including SOD2, prevent oxidative modification of calcium handling proteins such as the sarcoplasmic endoplasmic calcium ATPase (SERCA2A) and other cellular indices of calcium dyshomeostasis (French et al., 2006; French et al., 2008).

Finally, while exogenous antioxidant compounds (e.g., high doses of vitamin E, vitamin C, glutathione, etc.) are often beneficial in limiting ischemic injury in the heart, they can mitigate the redox-sensitive adaptive response to exercise (Ristow et al., 2009). In this regard, an important investigation by the Powers lab combined exercise and an exogenous antioxidant cocktail ( $\alpha$ -tocopherol, selenium, ascorbic acid, beta-carotene, CoQ10, N-acetyl cystine, and catechin) as a means of investigating the potentially additive effect against an ischemic insult. Findings revealed that exercise and antioxidants provided similar levels of independent cardioprotection, but that the combined dose of exercise and antioxidants did not potentiate the cardioprotective response (Hamilton et al., 2003). Further research is needed to resolve how long-term antioxidant dosing may alter exercise training outcomes, but as a preliminary conclusion it seems plausible that the exercised heart does not require the addition of antioxidant supplementation to bolster the cardioprotected phenotype.

#### Mitochondrial Proteins & Function

As described in the previous section, exercise induced cardiac preconditioning improves mitochondrial function during IR as a result of improved antioxidant status and calcium control (Bowles et al., 1992; French et al., 2006; Marcil et al., 2006; Starnes et al., 2007; French et al., 2008; Magalhaes et al., 2014). Subpopulations of subsarcolemmal and intermyofibrillar mitochondria exhibit distinct alterations following exercise, although more research is needed to fully understand these differences and functional relationship during IR (Kavazis et al., 2009). Both subpopulations from exercised hearts have decreased mPTP opening and decreased functional impairment

during IR compared to the sedentary condition (Kavazis et al., 2008; Lee et al., 2012). In addition to elevated mitochondrial antioxidants previously discussed, such as SOD2, a decrease in the enzyme monoamine oxidase (MAO), which catalyzes amine oxidation reactions and the production of reactive oxygen species (ROS), appears to be another mechanism by which exercise protects the heart from IR (Kavazis et al., 2009; Deshwal et al., 2017). Collectively, exercise prior to IR protects against mitochondrial release of ROS, activation of apoptotic signaling molecules, and mitochondrial damage and functional impairment (Kavazis et al., 2008; Kavazis et al., 2009; Lee et al., 2012).

#### Nitric Oxide

Nitric oxide appears to be important for exercise-induced cardioprotection against IR induced arrhythmias and injury (Hajnal et al., 2005; Akita et al., 2007). The function and synthesis of NO has been reviewed extensively elsewhere (Suhr et al., 2013; Wu et al., 2018). The exercise mediated cardioprotective effects of NO include angiogenesis and ventricular remodeling after myocardial infarction with protective cellular signaling through opening of ATP-sensitive potassium channels (K<sub>ATP</sub>) channels and a reduction in ROS (Wu et al., 2018). Nitric oxide also improves post-ischemic blood flow and contractile function (Pabla et al., 1995; Pabla et al., 1996). Nitric oxide production is enhanced following exercise training by increased activation and upregulation of eNOS (Fukai et al., 2000; Dao et al., 2011; Yang et al., 2014). Inducible nitric oxide synthase (iNOS) may also be involved in exercise mediated increases in NO, with some reports indicating that increases in eNOS following exercise training mediate elevated iNOS expression, which is required for exercise induced cardioprotection (Akita et al., 2007). However, not all studies have reported increases in iNOS (McGinnis et al., 2015). Additionally, as an inflammatory mediator, is likely not sustainable for a long period of time and some studies have indicated it is not essential for exercise induced cardiac preconditioning (Wilson et al., 2004; Quindry et al., 2010b; McGinnis et al., 2015). Activation of eNOS occurs via sympathetic nervous system activation and specifically β3-adrenergic receptors (Calvert et al., 2011). Elevated ROS and increased hydrogen peroxide also contribute to exercise mediated elevation in eNOS (Lauer et al., 2005; Akita et al., 2007). Although increases in antioxidant enzymes, such as genetic overexpression of catalase, result in diminished eNOS, catalase has not been consistently shown to increase following exercise training and eNOS remains increased following weeks of exercise training, indicating that any potential upregulation in antioxidant enzymes that might occur with exercise training are not inhibiting eNOS expression (Lauer et al., 2005; Dao et al., 2011; Yang et al., 2014). However, while exercise results in elevated eNOS in the myocardium and large arteries, this may not be the case in all tissues, as high catalase activity in venous tissue blocks exercise-induced increases in eNOS expression (Dao et al., 2011). Exercise-induced elevation of eNOS increases extracellular SOD, which then aids in maintaining NO levels by limiting superoxide mediated degradation of NO (Fukai et al., 2000).

#### Chaperone proteins and cardioprotection

An interesting facet of exercise and cardiac preconditioning is the lack of evidence for heat shock proteins (HSPs) as an essential mediator. Indeed, HPS expression is central to many forms of non-exercise preconditioning against ischemic insults, and exercise is well known to up-regulate HSPs, namely HSP72 (Demirel et al., 2003; Quindry et al., 2007). Nonetheless, experimental models have explored this topic using nonreductionist methods – performing treadmill exercise in a cold environment where the normal elevation in core temperatures is ~1.5-3 °C. In these investigations, mitigation in HSP72 expression appeared to have no bearing on various observations of cardioprotection against ischemic insults (Hamilton et al., 2001; Demirel et al., 2003; Quindry et al., 2007). Moreover, HSP family proteins have also been investigated and observed to be completely dissociated from corresponding observations of exercise-induced cardioprotection against ischemic injury (Hamilton et al., 2001).

It should be acknowledged, however, that these experimental approaches do not preclude the potential importance of HSPs in exercise-induced cardioprotection. Indeed, redundant mechanisms of protection highlighted within may mask the role of HPSs in the cardioprotected phenotype. One intriguing possibility is that HSPs may be playing a tissue specific role within the heart, and the current methodological design are insufficient to delineate the importance of HSPs within ventricular tissue versus an independent role in perfusion vessels. In support, an interesting study by Melling et al. (2007) revealed through confocal microscopy that HSP overexpression in the exercised heart was localized to the vasculature. Accordingly, the possibility remains that HSPs could yet prove essential to exercise preconditioning in the heart and/ or preserve vascular function during an ischemic challenge, although additional research is needed to confirm or refute this hypothesis. The reader is directed to a recent comprehensive review of the regulation of chaperone proteins as it pertains to cardiac preconditioning against IR (Penna et al., 2018).

#### Renin Angiotensin System

The renin angiotensin system (RAS) is comprised of two opposing arms: 1) the 'classical' arm, comprised of the angiotensin converting enzyme (ACE), and angiotensin I and angiotensin II receptors, and 2) 'counter regulatory arm' comprised of ACE2, the hepapeptide Ang-(1-7), and the Mas receptor (Ferrario et al., 1998; Donoghue et al., 2000; Nunes-Silva et al., 2017). Unlike the 'classical' arm, the ACE2/Mas receptor arm mediates anti-inflammatory, vasodilatory, and cardioprotective effects (Ferrario et al., 1998; Nunes-Silva et al., 2017). Exercise downregulates the classical RAS and upregulates the counter-regulatory RAS arm (Nunes-Silva et al., 2017; Magalhaes et al., 2020). More research is required to uncover the role of the dual arms of the RAS in exercise-induced cardiac preconditioning.

#### Channel function and improved metabolic control

Early investigations of exercise-induced cardioprotection using rat models of treadmill running and isolated heart preps revealed that calcium handling is significantly improved in exercised hearts (Bowles and Starnes, 1994). Indeed, parallel investigations of rats using isolated perfused working heart preparations from this group clearly indicate that improved calcium handling is inseparable from preservation of electron transport and tissue-level metabolism in exercised hearts (Bowles et al., 1992; Starnes and Bowles, 1995).

Roughly a decade later, it was discovered that  $K_{ATP}$  were also central to various forms of cardiac preconditioning of exercised hearts. Located on the sarcolemma (sarc  $K_{ATP}$ ) and the inner mitochondrial membrane (mito  $K_{ATP}$ ), the precise role of these putative mediators of cardioprotection is still debated (Light et al., 2001; Gross and Peart, 2003). Nonetheless, early findings in exercised hearts indicated that both sarc  $K_{ATP}$  and mito  $K_{ATP}$  are central to the exercised phenotype (Brown et al., 2005b; Johnson et al., 2006; Chicco et al., 2007). Interestingly, the only observations of sex-dependent differences in exerciseinduced cardioprotection appear to relate to  $K_{ATP}$  channel subunit expression, where hearts from exercised females appear to over-express the Kir6.2 subunit to a different level than males

#### (Johnson et al., 2006; Chicco et al., 2007).

While observations of variable K<sub>ATP</sub> channel sub-unit expression in exercised hearts were also linked to comparable sex-dependent differences in infarct sparing, it appears that hearts from exercised males are nonetheless protected via K<sub>ATP</sub> channel activation (Johnson et al., 2006; Chicco et al., 2007). To this end, and as mentioned earlier, the mechanisms of exercise-induced cardioprotection do not necessarily extend to all aspects of IR injury. Accordingly, an iterative series of experiments were performed where a common exercise regimen of treadmill running was applied against a variable duration of ischemic infarction as a means of evaluating the role of sarc  $K_{ATP}$  and mito  $K_{ATP}$  channels for preventing ventricular dysrhythmias and myocardial infarction. The shorter duration infarction study indicated that mito KATP channels were essential in preventing ventricular dysrhythmias, whereas the sarc K<sub>ATP</sub> channel was essential for preventing post-infarct tissue necrosis and apoptotic processes (Quindry et al., 2010a; Quindry et al., 2012). These integrated findings are also important in that the ventricular dysrhythmia sparing effects observed following the shorter duration ischemic event were no longer present when the ischemic time was increased by 150% (Quindry et al., 2010a; Quindry et al., 2012). The interpretation of these findings is that while exercise robustly protects against the evolving insult of an IR challenge, the phenotype is providing a time-dependent tolerance that has finite limits.

#### Autocrine and paracrine mediated protection

Perhaps most important to the sub-field of exercise-induced cardioprotection is the notion that exercise preconditioning provides a pragmatic scientific lens through which a druggable counter therapy to mitigate IR injury could be discovered. While there are multiple approaches to condition an organ through exogenous means, receptor-mediated approaches are among the first to be considered. Accordingly, a number of groups interested in exercise-induced cardioprotection have asked the question of whether the redundant protection afforded to exercised hearts might include receptor mediated processes. Indeed, this rationale might indicate tissue-to-tissue cross talk confers cardioprotection through circulating factors.

Given the parallel roles of exogenous opioids as a temporary approach to pharmacologic cardioprotection (Gross and Gross, 2007), and the reputed fact that aerobic exercise sometimes produces an endorphin response, it was speculated that some common mechanisms may mediate exercise-induced cardioprotection (Schultz et al., 1997; Karck et al., 2001; Schultz and Gross, 2001; Maslov et al., 2009). Opioid peptides increase following an acute bout of exercise (Kraemer et al., 1985; Goldfarb et al., 1987), supporting the rationale that endogenous opioids could have a role in exercise-induced cardioprotection. Endogenous opioids include three major families: β-endorphins, dynorphins, and enkephalins, as well as at least three major opioid receptor subtypes,  $(\mu, \delta, \kappa)$ , reviewed extensively elsewhere (van den Brink et al., 2003). Opioid peptides are released from various tissues, including the heart and skeletal muscle, with transcript expression of both opioid precursors and receptor subtypes increased in cardiac tissue following exercise (Denning et al., 2008; Dickson et al., 2008; Miller et al., 2015; Geng et al., 2017). Further, administration of a selective delta-opioid receptor agonist prior to exercise blunted the cardioprotective effect against tissue necrosis, suggesting the delta opioid receptor is involved in exercise induced cardioprotection (Miller et al., 2015). Some evidence also suggests that the kappa opioid receptor may also be involved in exercise induced cardioprotection, however not all studies have reported positive findings (Borges et al., 2014; Geng et al., 2017).

Although not definitive, it is most likely that opioid peptides

(leu-enkephalin) released from the heart are mediating the cardioprotective effect (Schultz et al., 1997; Maslov et al., 2009; Miller et al., 2015). However, delta opioid receptors are located only on intrinsic cardiac adrenergic (ICA) cells within the myocardium, and the cardioprotective effect of delta opioid receptor activation appears to be mediated by the release of epinephrine and calcitonin gene related peptide from ICA cells acting upon the β2-adrenergic and calcitonin generelated peptide (CGRP) receptors in cardiomyocytes (Huang et al., 2007; Huang et al., 2009). However, direct evidence that CGRP is released from ICA cells in response to exercisestimulated opioid release is yet to be confirmed. Additionally, cardioprotection afforded by circulating factors following exercise resulted in loss of protection against tissue necrosis when a non-selective opioid antagonist was administered, indicating circulating opioid molecules may have a role in exercise induced cardioprotection (Michelsen et al., 2012). It may be that opioid-mediated cardioprotection involves both opioids directly produced in cardiac tissue as well as other tissues delivered via circulation and various opioid receptor subtypes, although these conclusions require confirmation (Michelsen et al., 2012; Miller et al., 2015).

In addition, more research is needed to identify the interaction between opioid peptides, receptors and downstream signaling processes involved in exercise induced cardioprotection. To this end, while it cannot be ruled out that endorphins responsible for the often described "runner's high" may be cardioprotective, these factors are almost certainly not produced by the modest amounts of moderate intensity exercise observed to robustly protect against ischemic insults. This tentative conclusion is highly valuable to clinical and health-fitness narratives where patients and clients at significant risk for a heart attack can be assured that the best available scientific evidence indicates that one does not need to be a highly involved aerobic athlete to develop a cardioprotected heart.

#### Cytokines & Myokines

Extending the discussion of circulating factors and receptormediated cardioprotection, it is worth considering the role of cytokines/myokines and exercise-induced cardioprotection. By way of a brief review, inflammation is an important cellular response for survival and recovery. Yet for all the adaptive benefits of an acute inflammatory response of moderate intensity, it cannot be overlooked that dampening activation of inflammatory pathways in response to IR results in improved outcomes in terms of cell survival and function (Dawn et al., 2004). Central to this line of research is the molecule, interleukin-6 (IL-6). While IL-6 is traditionally held to be a potent cytokine, promoting inflammatory responses with significant collateral damage to healthy tissues, more recent evidence indicates that IL-6 may also have anti-inflammatory roles. While the full understanding of a dual functioning IL-6 is still being resolved, it appears that its role is contextual to the cytokine milieu. Thus, following a pathological stressor such as IR, the cytokine profile, which includes tumor necrosis factor alpha (TNF $\alpha$ ), results in IL-6 serving a deleterious role. However, following an acute bout of exercise, IL-6 is released in vesicles from contracting skeletal muscle, serving as an anti-inflammatory myokine, producing beneficial adaptative responses (Pedersen, 2009; Karstoft and Pedersen, 2016).

While not extensively examined in the context of exerciseinduced cardioprotection, one investigation provides preliminary insights. Using a whole-body IL-6 knockout mouse model of exercise and surgical LAD ligation, it was observed that IL-6 may be an important mediator of exercise preconditioning against both ventricular dysrhythmias and tissue death. Interestingly, the short-term exercise model resulted in over-expression of both soluble (circulating) and insoluble IL-6 receptors in the myocardium (McGinnis et al., 2015). This finding was interpreted to suggest that the benefits linking IL-6 function to exercise preconditioning may also confer cytoprotection to other tissues throughout the body through co-activation of IL-6 family receptor sub-types.

#### Exosomes

Recent studies suggest that exosomes, which transport various signaling molecules such as miRNAs, proteins, and lipids, may confer cardioprotective signals (Record et al., 2014; Vicencio et al., 2015; Hou et al., 2019). Some evidence suggests that exercise may confer protection against IR by increasing the circulating exosome concentration (Whitham et al., 2018). Indeed, exosomes isolated from plasma following exercise decreased infarct size and improved cardiac function following an experimental model of IR, although no differences in exosome number were found following exercise intervention (Hou et al., 2019). Specifically, exosomal miR-342-5p has been found to contribute to exercise derived cardioprotection by inhibiting apoptosis and promoting cell survival (Hou et al., 2019). Although most cell types secrete exosomes, it seems that vascular endothelial cells may be primarily responsible for increases in exosome miR-342-5p, which evidence indicates that flow-induced shear stress that occurs during exercise, acts as the signal (Hou et al., 2019). Additionally, it appears that cardiomyocytes have the ability to take up vascular endothelium derived exosomes, although others have proposed ligand-receptor mediated signaling (Vicencio et al., 2015; Hou et al., 2019). Exosome mediated cardioprotection has been demonstrated to involve HSP70 in models of nonstressed exosome (Vicencio et al., 2015). Cardiac upregulation of HSP70 has been found to not be essential for exercise induced cardioprotection (Hamilton et al., 2001; Quindry et al., 2007). It could be that protective pathways are activated in cardiomyocytes by exosomal HSP70 (Vicencio et al., 2015), opposed to upregulation within the cardiomyocyte. However, whether exercise cardioprotection involves exosomal associated HSP70 is currently unknown. Exosomes appear to be involved in various forms of cardiac preconditioning, such as remote ischemic preconditioning (Giricz et al., 2014; Vicencio et al., 2015). Future work in this exciting field to uncover mechanisms of exosome-derived cardioprotection are warranted.

#### **Evidence from human studies**

Evidence supporting exercise induced preconditioning in humans has been recently reviewed elsewhere (Thijssen et al., 2018) and will be briefly summarized here. Retrospective studies indicate improved survival rates for individuals with greater physical activity levels prior to an acute myocardial infarction and percutaneous transluminal coronary angioplasty (Abete et al., 2001; Rengo et al., 2007). Additionally, physical activity prior to coronary artery bypass graft improved outcomes by increasing survival rate (Rengo et al., 2010).

Human studies suggest a protective effect of previous exercise on angina and ST segment depression during a subsequent exercise session, referred to as 'warm-up angina'. It appears an initial stress test can decreased angina, later onset of ST segment depression, and decreased peak ST segment depression during a second stress test, lending support for the rationale that a single exercise session is protective against future myocardial ischemia in humans with angina (Lalonde et al., 2015). In contrast, the relatively moderate dose of exercise used in the majority of animal-based studies (without coronary artery disease) does not require the myocardium to become ischemic. However, mild to moderate exercise below the ischemic threshold was found to increase exercise duration and postpone angina onset, although this was related to decreased heart rate and blood pressure reactivity, so when compared to rate pressure product the angina onset was unchanged by moderate exercise, and only improved with relatively intense ischemia in the first exercise session (Bogaty et al., 2003). Still, existing evidence suggests that exercise preconditioning occurs in humans.

#### **Conclusions and future directions**

The field of exercise preconditioning continues to expand with important implications for both scientific exploration and clinical application. Due to the sustainable nature, effectiveness across the sexes and age groups, and evidence of protection in human clinical populations, more research and efforts for clinical application are needed. Although currently supported by human studies, additional work to clarify the specific exercise requirements and potential of exercise as a preconditioning stimulus prior to planned cardiac surgical procedures is warranted. Additionally, many preliminary findings detailed herein serve as a scientific rationale for the successful reverse engineering of cardioprotective counter therapies against IR injury. Perhaps just as important, exercise programming should be included in tandem with pharmacological prescriptions to treat and prevent cardiovascular and metabolic diseases. To this end, the reader is directed to the American College of Sports Medicine's Exercise is Medicine platform (https:// exerciseismedicine.org). Exercise is Medicine encompasses a wealth of refereed research detailing the cooperative interface between an exercise lifestyle and modern personalize medicine. Indeed, the tenets of Exercise is Medicine extend well beyond the current topic of exercise-induced cardiac preconditioning. In this regard however, the sub-field of exercise preconditioning is among the more scientifically progressive research lines, and it is tempting to speculate that many of the cytoprotective mechanisms already identified in the exercised heart are also responsible for the preconditioning effects descriptively observed in other organ systems throughout the body.

#### **Conflict of Interest Statement:**

The Authors declare that they have no conflicts of interest.

#### References

- Abete P, Ferrara N, Cacciatore F, Sagnelli E, Manzi M, Carnovale V, Calabrese C, de Santis D, Testa G, Longobardi G, Napoli C, Rengo F (2001) High level of physical activity preserves the cardioprotective effect of preinfarction angina in elderly patients. J Am Coll Cardiol 38:1357-1365.
- Akita Y, Otani H, Matsuhisa S, Kyoi S, Enoki C, Hattori R, Imamura H, Kamihata H, Kimura Y, Iwasaka T (2007) Exercise-induced activation of cardiac sympathetic nerve triggers cardioprotection via redox-sensitive activation of eNOS and upregulation of iNOS. Am J Physiol Heart Circ Physiol 292:H2051-2059.
- Bogaty P, Poirier P, Boyer L, Jobin J, Dagenais GR (2003) What induces the warm-up ischemia/angina phenomenon: exercise or myocardial ischemia? Circulation 107:1858-1863.
- Bolli R, Marban E (1999) Molecular and cellular mechanisms of myocardial stunning. Physiol Rev 79:609-634.
- Borges JP, Verdoorn KS, Daliry A, Powers SK, Ortenzi VH, Fortunato RS, Tibirica E, Lessa MA (2014) Delta opioid receptors: the link between exercise and cardioprotection. PLoS One 9:e113541.
- Bowles DK, Starnes JW (1994) Exercise training improves metabolic response after ischemia in isolated working rat heart. J Appl Physiol (1985) 76:1608-1614.
- Bowles DK, Farrar RP, Starnes JW (1992) Exercise training

improves cardiac function after ischemia in the isolated, working rat heart. Am J Physiol 263:H804-809.

- Brown DA, Moore RL (2007) Perspectives in innate and acquired cardioprotection: cardioprotection acquired through exercise. J Appl Physiol (1985) 103:1894-1899.
- Brown DA, Lynch JM, Armstrong CJ, Caruso NM, Ehlers LB, Johnson MS, Moore RL (2005a) Susceptibility of the heart to ischaemia-reperfusion injury and exerciseinduced cardioprotection are sex-dependent in the rat. J Physiol 564:619-630.
- Brown DA, Chicco AJ, Jew KN, Johnson MS, Lynch JM, Watson PA, Moore RL (2005b) Cardioprotection afforded by chronic exercise is mediated by the sarcolemmal, and not the mitochondrial, isoform of the KATP channel in the rat. J Physiol 569:913-924.
- Buja LM (2005) Myocardial ischemia and reperfusion injury. Cardiovasc Pathol 14:170-175.
- Calvert JW, Condit ME, Aragon JP, Nicholson CK, Moody BF, Hood RL, Sindler AL, Gundewar S, Seals DR, Barouch LA, Lefer DJ (2011) Exercise protects against myocardial ischemia-reperfusion injury via stimulation of beta(3)adrenergic receptors and increased nitric oxide signaling: role of nitrite and nitrosothiols. Circ Res 108:1448-1458.
- Chicco AJ, Johnson MS, Armstrong CJ, Lynch JM, Gardner RT, Fasen GS, Gillenwater CP, Moore RL (2007) Sex-specific and exercise-acquired cardioprotection is abolished by sarcolemmal KATP channel blockade in the rat heart. Am J Physiol Heart Circ Physiol 292:H2432-2437.
- Chowdhury MA, Sholl HK, Sharrett MS, Haller ST, Cooper CC, Gupta R, Liu LC (2019) Exercise and Cardioprotection: A Natural Defense Against Lethal Myocardial Ischemia-Reperfusion Injury and Potential Guide to Cardiovascular Prophylaxis. J Cardiovasc Pharmacol Ther 24:18-30.
- Criswell D, Powers S, Dodd S, Lawler J, Edwards W, Renshler K, Grinton S (1993) High intensity training-induced changes in skeletal muscle antioxidant enzyme activity. Med Sci Sports Exerc 25:1135-1140.
- Curtis MJ, Walker MJ (1988) Quantification of arrhythmias using scoring systems: an examination of seven scores in an in vivo model of regional myocardial ischaemia. Cardiovasc Res 22:656-665.
- Dao VT, Floeren M, Kumpf S, Both C, Peter B, Balz V, Suvorava T, Kojda G (2011) Catalase activity prevents exercise-induced up-regulation of vasoprotective proteins in venous tissue. J Cell Mol Med 15:2326-2334.
- Dawn B, Xuan YT, Guo Y, Rezazadeh A, Stein AB, Hunt G, Wu WJ, Tan W, Bolli R (2004) IL-6 plays an obligatory role in late preconditioning via JAK-STAT signaling and upregulation of iNOS and COX-2. Cardiovasc Res 64:61-71.
- Demirel HA, Hamilton KL, Shanely RA, Tumer N, Koroly MJ, Powers SK (2003) Age and attenuation of exerciseinduced myocardial HSP72 accumulation. Am J Physiol Heart Circ Physiol 285:H1609-1615.
- Demirel HA, Powers SK, Zergeroglu MA, Shanely RA, Hamilton K, Coombes J, Naito H (2001) Short-term exercise improves myocardial tolerance to in vivo ischemia-reperfusion in the rat. J Appl Physiol (1985) 91:2205-2212.
- Denning GM, Ackermann LW, Barna TJ, Armstrong JG, Stoll LL, Weintraub NL, Dickson EW (2008) Proenkephalin expression and enkephalin release are widely observed in non-neuronal tissues. Peptides 29:83-92.
- Deshwal S, Di Sante M, Di Lisa F, Kaludercic N (2017) Emerging role of monoamine oxidase as a therapeutic target for cardiovascular disease. Curr Opin Pharmacol 33:64-69.

- Dickson EW, Hogrefe CP, Ludwig PS, Ackermann LW, Stoll LL, Denning GM (2008) Exercise enhances myocardial ischemic tolerance via an opioid receptor-dependent mechanism. Am J Physiol Heart Circ Physiol 294:H402-408.
- Donoghue M, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, Donovan M, Woolf B, Robison K, Jeyaseelan R, Breitbart RE, Acton S (2000) A novel angiotensinconverting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9. Circ Res 87:E1-9.
- Feng R, Wang L, Li Z, Yang R, Liang Y, Sun Y, Yu Q, Ghartey-Kwansah G, Sun Y, Wu Y, Zhang W, Zhou X, Xu M, Bryant J, Yan G, Isaacs W, Ma J, Xu X (2019) A systematic comparison of exercise training protocols on animal models of cardiovascular capacity. Life Sci 217:128-140.
- Ferrario CM, Chappell MC, Dean RH, Iyer SN (1998) Novel angiotensin peptides regulate blood pressure, endothelial function, and natriuresis. J Am Soc Nephrol 9:1716-1722.
- Fiuza-Luces C, Santos-Lozano A, Joyner M, Carrera-Bastos P, Picazo O, Zugaza JL, Izquierdo M, Ruilope LM, Lucia A (2018) Exercise benefits in cardiovascular disease: beyond attenuation of traditional risk factors. Nat Rev Cardiol 15:731-743.
- Frasier CR, Sloan RC, Bostian PA, Gonzon MD, Kurowicki J, Lopresto SJ, Anderson EJ, Brown DA (2011) Short-term exercise preserves myocardial glutathione and decreases arrhythmias after thiol oxidation and ischemia in isolated rat hearts. J Appl Physiol (1985) 111:1751-1759.
- French JP, Hamilton KL, Quindry JC, Lee Y, Upchurch PA, Powers SK (2008) Exercise-induced protection against myocardial apoptosis and necrosis: MnSOD, calciumhandling proteins, and calpain. FASEB J 22:2862-2871.
- French JP, Quindry JC, Falk DJ, Staib JL, Lee Y, Wang KK, Powers SK (2006) Ischemia-reperfusion-induced calpain activation and SERCA2a degradation are attenuated by exercise training and calpain inhibition. Am J Physiol Heart Circ Physiol 290:H128-136.
- Fridovich I (1997) Superoxide anion radical (O2-.), superoxide dismutases, and related matters. J Biol Chem 272:18515-18517.
- Fukai T, Siegfried MR, Ushio-Fukai M, Cheng Y, Kojda G, Harrison DG (2000) Regulation of the vascular extracellular superoxide dismutase by nitric oxide and exercise training. J Clin Invest 105:1631-1639.
- Garber CE, Blissmer B, Deschenes MR, Franklin BA, Lamonte MJ, Lee IM, Nieman DC, Swain DP, American College of Sports M (2011) American College of Sports Medicine position stand. Quantity and quality of exercise for developing and maintaining cardiorespiratory, musculoskeletal, and neuromotor fitness in apparently healthy adults: guidance for prescribing exercise. Med Sci Sports Exerc 43:1334-1359.
- Geng X, Zhao H, Zhang S, Li J, Tian F, Feng N, Fan R, Jia M, Guo H, Cheng L, Liu J, Chen W, Pei J (2017) kappaopioid receptor is involved in the cardioprotection induced by exercise training. PLoS One 12:e0170463.
- Gielen S, Laughlin MH, O'Conner C, Duncker DJ (2015) Exercise training in patients with heart disease: review of beneficial effects and clinical recommendations. Prog Cardiovasc Dis 57:347-355.
- Giricz Z, Varga ZV, Baranyai T, Sipos P, Paloczi K, Kittel A, Buzas EI, Ferdinandy P (2014) Cardioprotection by remote ischemic preconditioning of the rat heart is mediated by extracellular vesicles. J Mol Cell Cardiol 68:75-78.

- Goldfarb AH, Hatfield BD, Sforzo GA, Flynn MG (1987) Serum beta-endorphin levels during a graded exercise test to exhaustion. Med Sci Sports Exerc 19:78-82.
- Grill HP, Zweier JL, Kuppusamy P, Weisfeldt ML, Flaherty JT (1992) Direct measurement of myocardial free radical generation in an in vivo model: effects of postischemic reperfusion and treatment with human recombinant superoxide dismutase. J Am Coll Cardiol 20:1604-1611.
- Gross ER, Gross GJ (2007) Pharmacologic therapeutics for cardiac reperfusion injury. Expert Opinions in Emergency Drugs 12:367-388.
- Gross GJ, Peart JN (2003) KATP channels and myocardial preconditioning: an update. Am J Physiol Heart Circ Physiol 285:H921-930.
- Hajnal A, Nagy O, Litvai A, Papp J, Parratt JR, Vegh A (2005) Nitric oxide involvement in the delayed antiarrhythmic effect of treadmill exercise in dogs. Life Sci 77:1960-1971.
- Hamilton KL, Staib JL, Phillips T, Hess A, Lennon SL, Powers SK (2003) Exercise, antioxidants, and HSP72: protection against myocardial ischemia/reperfusion. Free Radic Biol Med 34:800-809.
- Hamilton KL, Powers SK, Sugiura T, Kim S, Lennon S, Tumer N, Mehta JL (2001) Short-term exercise training can improve myocardial tolerance to I/R without elevation in heat shock proteins. Am J Physiol Heart Circ Physiol 281:H1346-1352.
- Hou Z, Qin X, Hu Y, Zhang X, Li G, Wu J, Li J, Sha J, Chen J, Xia J, Wang L, Gao F (2019) Longterm Exercise-Derived Exosomal miR-342-5p. Circ Res 124:1386-1400.
- Huang MH, Wang HQ, Roeske WR, Birnbaum Y, Wu Y, Yang NP, Lin Y, Ye Y, McAdoo DJ, Hughes MG, Lick SD, Boor PJ, Lui CY, Uretsky BF (2007) Mediating deltaopioid-initiated heart protection via the beta2-adrenergic receptor: role of the intrinsic cardiac adrenergic cell. Am J Physiol Heart Circ Physiol 293:H376-384.
- Huang MH, Nguyen V, Wu Y, Rastogi S, Lui CY, Birnbaum Y, Wang HQ, Ware DL, Chauhan M, Garg N, Poh KK, Ye L, Omar AR, Tan HC, Uretsky BF, Fujise K (2009) Reducing ischaemia/reperfusion injury through deltaopioid-regulated intrinsic cardiac adrenergic cells: adrenopeptidergic co-signalling. Cardiovasc Res 84:452-460.
- Johnson MS, Moore RL, Brown DA (2006) Sex differences in myocardial infarct size are abolished by sarcolemmal KATP channel blockade in rat. Am J Physiol Heart Circ Physiol 290:H2644-2647.
- Karck M, Tanaka S, Bolling SF, Simon A, Su TP, Oeltgen PR, Haverich A (2001) Myocardial protection by ischemic preconditioning and delta-opioid receptor activation in the isolated working rat heart. J Thorac Cardiovasc Surg 122:986-992.
- Karstoft K, Pedersen BK (2016) Skeletal muscle as a gene regulatory endocrine organ. Curr Opin Clin Nutr Metab Care 19:270-275.
- Kavazis AN, McClung JM, Hood DA, Powers SK (2008) Exercise induces a cardiac mitochondrial phenotype that resists apoptotic stimuli. Am J Physiol Heart Circ Physiol 294:H928-935.
- Kavazis AN, Alvarez S, Talbert E, Lee Y, Powers SK (2009) Exercise training induces a cardioprotective phenotype and alterations in cardiac subsarcolemmal and intermyofibrillar mitochondrial proteins. Am J Physiol Heart Circ Physiol 297:H144-152.
- Kraemer WJ, Noble B, Culver B, Lewis RV (1985) Changes in plasma proenkephalin peptide F and catecholamine levels during graded exercise in men. Proc Natl Acad Sci U S A

82:6349-6351.

- Kupai K, Szabo R, Veszelka M, Awar AA, Torok S, Csonka A, Barath Z, Posa A, Varga C (2015) Consequences of exercising on ischemia-reperfusion injury in type 2 diabetic Goto-Kakizaki rat hearts: role of the HO/NOS system. Diabetol Metab Syndr 7:85.
- Lalonde F, Poirier P, Sylvestre MP, Arvisais D, Curnier D (2015) Exercise-induced ischemic preconditioning detected by sequential exercise stress tests: a meta-analysis. Eur J Prev Cardiol 22:100-112.
- Lauer N, Suvorava T, Ruther U, Jacob R, Meyer W, Harrison DG, Kojda G (2005) Critical involvement of hydrogen peroxide in exercise-induced up-regulation of endothelial NO synthase. Cardiovasc Res 65:254-262.
- Lawler JM, Powers SK, Hammeren J, Martin AD (1993) Oxygen cost of treadmill running in 24-month-old Fischer-344 rats. Med Sci Sports Exerc 25:1259-1264.
- Lee Y, Min K, Talbert EE, Kavazis AN, Smuder AJ, Willis WT, Powers SK (2012) Exercise protects cardiac mitochondria against ischemia-reperfusion injury. Med Sci Sports Exerc 44:397-405.
- Lefer DJ, Bolli R (2011) Development of an NIH consortium for preclinicAl AssESsment of CARdioprotective therapies (CAESAR): a paradigm shift in studies of infarct size limitation. J Cardiovasc Pharmacol Ther 16:332-339.
- Lennon SL, Quindry JC, French JP, Kim S, Mehta JL, Powers SK (2004a) Exercise and myocardial tolerance to ischaemia-reperfusion. Acta Physiol Scand 182:161-169.
- Lennon SL, Quindry J, Hamilton KL, French J, Staib J, Mehta JL, Powers SK (2004b) Loss of exercise-induced cardioprotection after cessation of exercise. J Appl Physiol (1985) 96:1299-1305.
- Light PE, Kanji HD, Fox JE, French RJ (2001) Distinct myoprotective roles of cardiac sarcolemmal and mitochondrial KATP channels during metabolic inhibition and recovery. Faseb J 15:2586-2594.
- Lightfoot JT, Leamy L, Pomp D, Turner MJ, Fodor AA, Knab A, Bowen RS, Ferguson D, Moore-Harrison T, Hamilton A (2010) Strain screen and haplotype association mapping of wheel running in inbred mouse strains. J Appl Physiol (1985) 109:623-634.
- Lund J, Hafstad AD, Boardman NT, Rossvoll L, Rolim NP, Ahmed MS, Florholmen G, Attramadal H, Wisloff U, Larsen TS, Aasum E (2015) Exercise training promotes cardioprotection through oxygen-sparing action in high fat-fed mice. Am J Physiol Heart Circ Physiol 308:H823-829.
- Magalhaes DM, Nunes-Silva A, Rocha GC, Vaz LN, de Faria MHS, Vieira ELM, Rocha NP, Simoes ESAC (2020) Two protocols of aerobic exercise modulate the counterregulatory axis of the renin-angiotensin system. Heliyon 6:e03208.
- Magalhaes J, Goncalves IO, Lumini-Oliveira J, Marques-Aleixo I, Passos E, Rocha-Rodrigues S, Machado NG, Moreira AC, Rizo D, Viscor G, Oliveira PJ, Torrella JR, Ascensao A (2014) Modulation of cardiac mitochondrial permeability transition and apoptotic signaling by endurance training and intermittent hypobaric hypoxia. Int J Cardiol 173:40-45.
- Mancardi D, Tullio F, Crisafulli A, Rastaldo R, Folino A, Penna C, Pagliaro P (2009) Omega 3 has a beneficial effect on ischemia/reperfusion injury, but cannot reverse the effect of stressful forced exercise. Nutr Metab Cardiovasc Dis 19:20-26.
- Marcil M, Bourduas K, Ascah A, Burelle Y (2006) Exercise training induces respiratory substrate-specific decrease in Ca2+-induced permeability transition pore opening

in heart mitochondria. Am J Physiol Heart Circ Physiol 290:H1549-1557.

- Marshall KD, Muller BN, Krenz M, Hanft LM, McDonald KS, Dellsperger KC, Emter CA (2013) Heart failure with preserved ejection fraction: chronic low-intensity interval exercise training preserves myocardial O2 balance and diastolic function. J Appl Physiol (1985) 114:131-147.
- Maslov LN, Lishmanov YB, Oeltgen PR, Barzakh EI, Krylatov AV, Govindaswami M, Brown SA (2009) Activation of peripheral delta2 opioid receptors increases cardiac tolerance to ischemia/reperfusion injury Involvement of protein kinase C, NO-synthase, KATP channels and the autonomic nervous system. Life Sci 84:657-663.
- McGinnis GR, Ballmann C, Peters B, Nanayakkara G, Roberts M, Amin R, Quindry JC (2015) Interleukin-6 mediates exercise preconditioning against myocardial ischemia reperfusion injury. Am J Physiol Heart Circ Physiol 308:H1423-1433.
- Melling CW, Thorp DB, Milne KJ, Krause MP, Noble EG (2007) Exercise-mediated regulation of Hsp70 expression following aerobic exercise training. Am J Physiol Heart Circ Physiol 293:H3692-3698.
- Michelsen MM, Stottrup NB, Schmidt MR, Lofgren B, Jensen RV, Tropak M, St-Michel EJ, Redington AN, Botker HE (2012) Exercise-induced cardioprotection is mediated by a bloodborne, transferable factor. Basic Res Cardiol 107:260.
- Miller LE, Hosick PA, Wrieden J, Hoyt E, Quindry JC (2012) Evaluation of arrhythmia scoring systems and exerciseinduced cardioprotection. Med Sci Sports Exerc 44:435-441.
- Miller LE, McGinnis GR, Peters BA, Ballmann CG, Nanayakkara G, Amin R, Quindry JC (2015) Involvement of the delta-opioid receptor in exercise-induced cardioprotection. Exp Physiol 100:410-421.
- Nunes-Silva A, Rocha GC, Magalhaes DM, Vaz LN, Salviano de Faria MH, Simoes ESAC (2017) Physical Exercise and ACE2-Angiotensin-(1-7)-Mas Receptor Axis of the Renin Angiotensin System. Protein Pept Lett 24:809-816.
- Pabla R, Buda AJ, Flynn DM, Salzberg DB, Lefer DJ (1995) Intracoronary nitric oxide improves postischemic coronary blood flow and myocardial contractile function. Am J Physiol 269:H1113-1121.
- Pabla R, Buda AJ, Flynn DM, Blesse SA, Shin AM, Curtis MJ, Lefer DJ (1996) Nitric oxide attenuates neutrophilmediated myocardial contractile dysfunction after ischemia and reperfusion. Circ Res 78:65-72.
- Paffenbarger RS, Jr., Wolf PA, Notkin J, Thorne MC (1966) Chronic disease in former college students. I. Early precursors of fatal coronary heart disease. Am J Epidemiol 83:314-328.
- Peart JN, Hoe LE, Gross GJ, Headrick JP (2011) Sustained ligand-activated preconditioning via delta-opioid receptors. The Journal of pharmacology and experimental therapeutics 336:274-281.
- Pedersen BK (2009) Edward F. Adolph distinguished lecture: muscle as an endocrine organ: IL-6 and other myokines. J Appl Physiol 107:1006-1014.
- Penna C, Alloatti G, Crisafulli A (2020) Mechanisms Involved in Cardioprotection Induced by Physical Exercise. Antioxid Redox Signal 32:1115-1134.
- Penna C, Sorge M, Femmino S, Pagliaro P, Brancaccio M (2018) Redox Aspects of Chaperones in Cardiac Function. Front Physiol 9:216.
- Pons S, Martin V, Portal L, Zini R, Morin D, Berdeaux A, Ghaleh B (2013) Regular treadmill exercise restores cardioprotective signaling pathways in obese mice independently from improvement in associated co-

- Powers SK, Smuder AJ, Kavazis AN, Quindry JC (2014) Mechanisms of exercise-induced cardioprotection. Physiology (Bethesda) 29:27-38.
- Powers SK, Demirel HA, Vincent HK, Coombes JS, Naito H, Hamilton KL, Shanely RA, Jessup J (1998) Exercise training improves myocardial tolerance to in vivo ischemia-reperfusion in the rat. Am J Physiol 275:R1468-1477.
- Quindry J, French J, Hamilton K, Lee Y, Mehta JL, Powers S (2005) Exercise training provides cardioprotection against ischemia-reperfusion induced apoptosis in young and old animals. Exp Gerontol 40:416-425.
- Quindry JC (2017) Exercise-Induced Cardioprotection and the Therapeutic Potential of RIPC. J Cardiovasc Pharmacol Ther 22:397-403.
- Quindry JC, Franklin BA (2018) Cardioprotective Exercise and Pharmacologic Interventions as Complementary Antidotes to Cardiovascular Disease. Exerc Sport Sci Rev 46:5-17.
- Quindry JC, Schreiber L, Hosick P, Wrieden J, Irwin JM, Hoyt E (2010a) Mitochondrial KATP channel inhibition blunts arrhythmia protection in ischemic exercised hearts. Am J Physiol Heart Circ Physiol 299:H175-183.
- Quindry JC, French J, Hamilton KL, Lee Y, Selsby J, Powers S (2010b) Exercise does not increase cyclooxygenase-2 myocardial levels in young or senescent hearts. J Physiol Sci 60:181-186.
- Quindry JC, Hamilton KL, French JP, Lee Y, Murlasits Z, Tumer N, Powers SK (2007) Exercise-induced HSP-72 elevation and cardioprotection against infarct and apoptosis. J Appl Physiol (1985) 103:1056-1062.
- Quindry JC, Miller L, McGinnis G, Kliszczewicz B, Irwin JM, Landram M, Urbiztondo Z, Nanayakkara G, Amin R (2012) Ischemia reperfusion injury, KATP channels, and exercise-induced cardioprotection against apoptosis. J Appl Physiol (1985) 113:498-506.
- Record M, Poirot M, Silvente-Poirot S (2014) Emerging concepts on the role of exosomes in lipid metabolic diseases. Biochimie 96:67-74.
- Rengo G, Galasso G, Vitale DF, Furgi G, Zincarelli C, Golino L, Femminella GD, Piscione F, Rengo F, Leosco D (2010) An active lifestyle prior to coronary surgery is associated with improved survival in elderly patients. J Gerontol A Biol Sci Med Sci 65:758-763.
- Rengo G, Galasso G, Piscione F, Golino L, Fortunato F, Zincarelli C, Cassese S, Abete P, Chiariello M, Rengo F, Leosco D (2007) An active lifestyle improves outcome of primary angioplasty in elderly patients with acute myocardial infarction. Am Heart J 154:352-360.
- Riebe D, Ehrman JK, Liguori G, Magal M (2018) ACSM's guidelines for exercise testing and prescription, Tenth edition. Edition. Philadelphia: Wolters Kluwer.
- Ristow M, Zarse K, Oberbach A, Kloting N, Birringer M, Kiehntopf M, Stumvoll M, Kahn CR, Bluher M (2009) Antioxidants prevent health-promoting effects of physical exercise in humans. Proc Natl Acad Sci U S A 106:8665-8670.
- Schultz JE, Gross GJ (2001) Opioids and cardioprotection. Pharmacol Ther 89:123-137.
- Schultz JJ, Hsu AK, Gross GJ (1997) Ischemic preconditioning and morphine-induced cardioprotection involve the delta (delta)-opioid receptor in the intact rat heart. J Mol Cell Cardiol 29:2187-2195.
- Soufi FG, Saber MM, Ghiassie R, Alipour M (2011) Role of 12-week resistance training in preserving the heart against ischemia-reperfusion-induced injury. Cardiol J 18:140-145.

Starnes JW, Bowles DK (1995) Role of exercise in the cause

and prevention of cardiac dysfunction. Exercise and sport sciences reviews 23:349-373.

- Starnes JW, Taylor RP (2007) Exercise-induced cardioprotection: endogenous mechanisms. Med Sci Sports Exerc 39:1537-1543.
- Starnes JW, Taylor RP, Park Y (2003) Exercise improves postischemic function in aging hearts. Am J Physiol Heart Circ Physiol 285:H347-351.
- Starnes JW, Taylor RP, Ciccolo JT (2005) Habitual lowintensity exercise does not protect against myocardial dysfunction after ischemia in rats. Eur J Cardiovasc Prev Rehabil 12:169-174.
- Starnes JW, Barnes BD, Olsen ME (2007) Exercise training decreases rat heart mitochondria free radical generation but does not prevent Ca2+-induced dysfunction. J Appl Physiol (1985) 102:1793-1798.
- Suhr F, Gehlert S, Grau M, Bloch W (2013) Skeletal muscle function during exercise-fine-tuning of diverse subsystems by nitric oxide. Int J Mol Sci 14:7109-7139.
- Taylor RP, Olsen ME, Starnes JW (2007) Improved postischemic function following acute exercise is not mediated by nitric oxide synthase in the rat heart. Am J Physiol Heart Circ Physiol 292:H601-607.
- Thijssen DHJ, Redington A, George KP, Hopman MTE, Jones H (2018) Association of Exercise Preconditioning With Immediate Cardioprotection: A Review. JAMA Cardiol 3:169-176.
- van den Brink OW, Delbridge LM, Rosenfeldt FL, Penny D, Esmore DS, Quick D, Kaye DM, Pepe S (2003) Endogenous cardiac opioids: enkephalins in adaptation and protection of the heart. Heart Lung Circ 12:178-187.
- Vicencio JM, Yellon DM, Sivaraman V, Das D, Boi-Doku C, Arjun S, Zheng Y, Riquelme JA, Kearney J, Sharma V, Multhoff G, Hall AR, Davidson SM (2015) Plasma exosomes protect the myocardium from ischemiareperfusion injury. J Am Coll Cardiol 65:1525-1536.

- Virani SS et al. (2020) Heart Disease and Stroke Statistics-2020 Update: A Report From the American Heart Association. Circulation 141:e139-e596.
- Whitham M, Parker BL, Friedrichsen M, Hingst JR, Hjorth M, Hughes WE, Egan CL, Cron L, Watt KI, Kuchel RP, Jayasooriah N, Estevez E, Petzold T, Suter CM, Gregorevic P, Kiens B, Richter EA, James DE, Wojtaszewski JFP, Febbraio MA (2018) Extracellular Vesicles Provide a Means for Tissue Crosstalk during Exercise. Cell Metab 27:237-251 e234.
- Wilson EM, Diwan A, Spinale FG, Mann DL (2004) Duality of innate stress responses in cardiac injury, repair, and remodeling. J Mol Cell Cardiol 37:801-811.
- Wojcik B, Knapp M, Gorski J (2018) Non-ischemic heart preconditioning. J Physiol Pharmacol 69.
- Wu D, Hu Q, Zhu D (2018) An Update on Hydrogen Sulfide and Nitric Oxide Interactions in the Cardiovascular System. Oxid Med Cell Longev 2018:4579140.
- Yamashita N, Hoshida S, Otsu K, Asahi M, Kuzuya T, Hori M (1999) Exercise provides direct biphasic cardioprotection via manganese superoxide dismutase activation. J Exp Med 189:1699-1706.
- Yang L, Jia Z, Yang L, Zhu M, Zhang J, Liu J, Wu P, Tian W, Li J, Qi Z, Tang X (2014) Exercise protects against chronic beta-adrenergic remodeling of the heart by activation of endothelial nitric oxide synthase. PLoS One 9:e96892.
- Zweier JL (1988) Measurement of superoxide-derived free radicals in the reperfused heart. Evidence for a free radical mechanism of reperfusion injury. J Biol Chem 263:1353-1357.