

REVIEW ARTICLE | OPEN ACCESS

Hypoxia in the CNS: a focus on multiple sclerosis

Sebok K. Halder¹, Richard Milner¹

Abstract Multiple sclerosis (MS) is the most common neurological disease affecting the young-middle age population in the Western hemisphere. Pathologically, it is a chronic inflammatory disease in which autoreactive T lymphocytes cross the blood-brain barrier to destroy the myelin insulating sheaths around axons. While accumulating evidence suggests a potential role for hypoxia in disease pathogenesis, interestingly, recent studies in the mouse MS model experimental autoimmune encephalomyelitis (EAE) have demonstrated the protective effect of hypoxic conditioning, both in the form of pre-conditioning and more importantly, when used to treat pre-existing disease. In this review we summarize the findings of these studies with a focus on: (i) the molecular mechanisms underlying the protective effect of hypoxic conditioning, and (ii) the potential clinical implications of these findings.

Keywords: multiple schlerosis, hypoxia, blood vessels, inflammation, blood-brain barrier integrity, angiogenesis,

Introduction

1. Multiple sclerosis is a heterogenous disease

Multiple sclerosis (MS) is the most common neurological disease affecting the young and middle-aged in the Western hemisphere, with a prevalence of 1.4 in 1000 in North America and 1.1 in 1000 in Europe. It has a gender preference, being 2-3 times more common in women than men (Wingerchuk and Carter, 2014; Brownlee et al., 2017; Doshi and Chataway, 2017). At the clinical level, it presents with a broad spectrum of signs and symptoms including visual disturbance, loss of sensation, weakness, paralysis, and incontinence, as well as cognitive defects, which together, reflect disrupted neural transmission in the visual, motor, sensory, and autonomic pathways. At the pathological level, MS is a chronic inflammatory disease that leads to stripping of the insulating myelin sheath that wraps around axons, a process known as demyelination. If the axons are left unmyelinated for extended periods of time, axonal degeneration ultimately ensues, resulting in permanent disability (French-Constant, 1994; Lassmann, 1998; Compston and Coles, 2008). MS is a highly complex disease, with patients showing tremendous heterogeneity not just in the way their disease manifests, but also in how it progresses and responds to different medications (Wingerchuk and Carter, 2014; Brownlee et al., 2017; Doshi and Chataway, 2017).

To try and understand why so much variation exists between different MS patients, more than twenty years ago, a series of studies examined a large number of MS lesions at the histopathological level and this revealed some insightful findings. Interestingly, while all active MS lesions were characterized by a chronic inflammatory demyelinating process mediated by T lymphocytes and macrophages, the immunological and cell-specific structural components varied significantly between different patients. By careful analysis, four basic patterns were described (Lucchinetti et al., 1996; Lucchinetti et al., 1999, 2000). In the first (pattern I), the demyelinating lesion included inflammatory T cells and macrophages wrapped around blood vessels but showed relative preservation of myelin-producing cells (oligodendrocytes) and no antibody involvement (Brück et al., 1995; Lucchinetti et al., 1996; Gay et al., 1997; Babbe et al., 2000). Pattern II showed similar characteristics to pattern I but also included the accumulation of antibody and complement on degenerating myelin, suggesting the presence of an underlying antibodymediated pathogenic process (Prineas and Graham, 1981; Storch et al., 1998a; Storch et al., 1998b; Diaz-Villoslada et al., 1999; Genain et al., 1999). Pattern IV was unique in showing oligodendrocyte death within the focus of the demyelinating lesion, but no antibody deposition, suggesting that this type of lesion is caused by a defect in oligodendrocytes, i.e., is

¹San Diego Biomedical Research Institute, 10865 Road to the Cure, Suite 100, San Diego, CA 92121, USA.

Correspondence should be addressed to Dr. Richard Milner (rmilner@sdbri.org).

a primary oligodendrogliopathy (Lucchinetti et al., 1996). Pattern III was also highly distinct in that it showed little to no inflammation but was described as a "dying back" oligodendrogliopathy, whereby oligodendroglial proteins located at the distal end of oligodendrocyte processes, such as myelin associated glycoprotein (MAG) and cyclic nucleotide phosphodiesterase (CNPase) were completely absent, while myelin proteins concentrated at a more central position on oligodendrocytes (in the compact myelin), such as myelin basic protein (MBP) and proteolipid protein (PLP), remained intact (Itoyama et al., 1980; Ludwin and Johnson, 1981; Lucchinetti et al., 2000). With progression of this type of lesion, the destruction appeared to spread more centrally, leading ultimately to oligodendroglial apoptosis. Because of the progressive nature of type III lesion pathology, it was initially thought that this may represent an early evolutionary stage of all types of MS lesions (Lassmann, 2003; Barnett and Prineas, 2004; Barnett et al., 2009) but later studies that demonstrated the persistence of individual-based different types of MS lesion over time suggests that the type III lesion is a distinct entity in its own right (König et al., 2008; Metz et al., 2014). In light of the unique nature of this "dying back" oligodendrogliopathy, subsequent studies began to analyze whether this type of demyelinating lesion is also present in animal models of MS or in other human neurological diseases. This revealed that while it is not observed in the most widely used animal model of MS, experimental autoimmune encephalomyelitis (EAE), it is a characteristic feature of oligodendrocyte degeneration in the cuprizone demyelination model, in which the toxin cuprizone interferes with energy metabolism within oligodendrocytes, perhaps providing the first clue that deficient energy metabolism might be a potential cause of this "dying back" oligodendrogliopathy (Ludwin and Johnson, 1981). In other human diseases, distal oligodendrogliopathy was also demonstrated in some cases of virus-induced white matter inflammation, including herpes simplex and cytomegalovirus-induced encephalomyelitis (Rodriguez, 1992), as well as the often fatal condition, progressive multifocal leukoencephalopathy (PML) (Itoyama et al., 1982). Most interestingly, distal oligodendrogliopathy is also a robust and consistent feature of acute white matter stroke, raising the possibility that ischemic damage may be the primary trigger resulting in the dying back of oligodendrocytes in type III MS lesions (Aboul-Enein et al., 2003). Further analysis revealed that amongst the different types of MS lesions, only type III lesions show high expression levels of the master regulatory transcription factor, hypoxia-inducible factor (HIF)- 1α (Aboul-Enein et al., 2003). Taken together, these findings prompted the suggestion that hypoxia may play a central role in driving white matter inflammation and oligodendrocyte death in MS, at least in the type III subset of MS lesions.

2. Is hypoxia a contributory factor in the pathogenesis of demyelinating disease?

While MS is traditionally described as a chronic inflammatory disease of autoimmune origin in which autoreactive T lymphocytes cross the blood-brain barrier (BBB) to destroy myelin, the precise sequence of events that triggers the autoimmune attack are still unknown. Over the last thirty years, a growing number of studies have provided evidence supporting the idea that hypoxia may be an etiological factor in MS, either as the initial trigger or in maintaining the ongoing inflammatory attack. This was first suggested by the similarity between the dying back oligodendrogliopathy common to type III MS lesions and acute white matter stroke, suggesting that a hypoxia-driven pathogenic mechanism may underlie this type of MS lesion (Lucchinetti et al., 1999, 2000). This concept was supported by the finding that HIF-1 α expression was found in oligodendrocytes and other cell types in type III lesions

but not in other patterns of MS lesions (Aboul-Enein et al., 2003). Subsequent studies confirmed hypoxic-like injury in the brains of MS patients as well as implicating the inflammatory mediators, reactive oxygen species (ROS) and nitric oxide (NO) in mediating mitochondrial dysfunction (Redford et al., 1997a; Aboul-Enein and Lassmann, 2005). Other studies using microarray analysis demonstrated that MS tissue showed upregulation of genes involved in ischemic pre-conditioning, such as HIF-1 α and its target, vascular endothelial growth factor receptor (VEGFR)-1, as well as those involved in mediating neuroprotective responses against oxidative stress, while another described increased levels of nitric oxide synthetase (NOS), NO, and adjacent oligodendrocyte damage, as well as induction of heme oxygenase-1 (Graumann et al., 2003; Zeis et al., 2009). Furthermore, marked suppression of mitochondrial respiratory chain complexes was observed in degenerating axons in MS lesions (Dutta et al., 2006). Based on these observations, it was hypothesized that during progression of the demyelinating lesion, inflammatory cells release ROS and NO, which then inhibit mitochondrial function in demyelinated axons, resulting in an energy-deficient metabolic crisis. This situation was described as "virtual hypoxia," implying not a specific lack of oxygen per se, but rather an energy-deficient state caused by NO and ROS-induced mitochondrial dysfunction (Trapp and Stys, 2009). These findings have since been substantiated in recent studies and it is now widely accepted that "virtual hypoxia" is an important pathogenic component of the demyelinating lesion (Mahad et al., 2015).

In addition to the contribution of "virtual hypoxia" to MS pathogenesis, during the last decade, a series of studies examining oxygen levels both in the brains of the EAE animals and in living MS patients have definitively demonstrated that a state of true hypoxia, or oxygen deficiency, exists within demyelinating lesions. In the spinal cord of EAE rats, Davies et al. (2013) demonstrated robust hypoxia using implanted oxygen probes and pimonidazole labeling. Interestingly, the level of hypoxia closely correlated with the degree of neurological deficit, with spinal cord pO₂ levels falling from 35 mm Hg in control disease-free animals to an average of 20 mm Hg in animals with hindlimb paralysis. Of note, pO₂ levels returned to normal during clinical remission but spinal cord tissue became hypoxic again during clinical relapse. This study also made the important observation that pO_2 levels in spinal cord white matter were always lower than in gray matter, in keeping with the lower levels of vascularity and greater hypoxic vulnerability demonstrated in these regions (Hassler, 1966; Turnbull et al., 1966; Tveten, 1976; Koyanagi et al., 1993a, b; Halder et al., 2018b).

In follow-up studies, Desai et al. (2016) devised an approach to model the molecular events underlying the early development of an inflammatory demyelinating lesion by focal injection of lipopolysaccharide into the spinal cord. In this model, they detected transient hypoxia at an early timepoint of disease progression within the white-gray matter border and dorsal white column (watershed) areas of the spinal cord and found that this was associated with increased ROS and NO, which over time, led to marked oligodendrocyte loss and demyelination (Desai et al., 2016). These observations led the authors to suggest a model in which activation of innate immune cells within the inflamed spinal cord triggers transient hypoxia in vascular watershed areas, culminating in a state of energy deficiency that leads to oligodendrocyte death and demyelination.

These observations have been substantiated by several key studies from the Dunn laboratory using a variety of novel techniques to demonstrate true hypoxia in demyelinating lesions both in the central nervous system (CNS) of EAE animals and MS patients. By using susceptibility-weighted imaging (SWI), Table 1. A summary of the potential causes of hypoxia in MS.

The watershed concept: cerebrovascular anatomy	
- Demyelination typically occurs at terminal endpoints of arterioles where blood supply is weakest	Brownell and Hughes, 1962; Hassler, 1966; Turnbull, 1966; Tveten, 1976; Haider, 2016.
Cerebral hypoperfusion due to underlying vascular predisposition	Swank, 1983; Brooks, 1984
 Associated with cardiovascular risk factors: diabetes, dyslipidemia, smoking Overlap with migraine; thickening of arteriolar walls Impaired vasodilatory responses in MS patients Elevated endothelin-1 (vasoconstrictor) levels in MS patients 	Kappus, 2016; Bechtold, 2014; Giubilei, 2011; Hernan, 2001. Poyrazoglu, 2016; Wang, 2019; Yuksel, 2019. Ranadive, 2012; Marshall, 2014. Speciale, 2000; Haufschild, 2001.
Vascular inflammation: endothelial activation	
Unregulation of NUC close II and fibrin deposition in MC patients	Weksfield 1004
- Opregulation of MHC class II and librin deposition in MS patients	Wakeneid, 1994.
Systemic hypoxia	
 Chronic obstructive pulmonary disease (COPD) Obstructive sleep apnea Impaired cardiopulmonary function 	Yohannes, 2017; Russ, 2020. Bucks, 2003; Braley, 2012. Bandopadhyay, 2000.
"Virtual hypoxia"	
- Inflammatory cells release nitric oxide (NO) and reactive oxygen species (ROS), which inhibit mitochondrial function in demyelinated axons, resulting in energy-deficient crisis	Dutta, 2006; Trapp, 2009; Mahad, 2015.

an iron-sensitive magnetic resonance imaging (MRI) method, that is typically used to detect iron deposition in tissues, Nathoo et al. (2013) modified this technique to estimate the levels of deoxyhemoglobin in blood vessels. This showed that SWI lesions were present in the spinal cord and cerebellum of EAE mice but importantly, many of these lesions disappeared following perfusion (removal of blood), indicating that these lesions represent deoxyhemoglobin (i.e., poorly oxygenated blood) within blood vessels (Nathoo et al., 2013). In a followup study, the same group showed that by increasing the level of inspired oxygen (from 30 to 100%), the SWI lesions in EAE mice largely disappeared (Nathoo et al., 2015). This implies that the majority of SWI lesions were not due to iron deposition within tissue per se, but rather due to increased levels of deoxyhemoglobin within blood vessels, because when oxygen supply was increased, the oxygen deficit within blood vessels disappeared. This reinforces the concept that a state of true oxygen deficiency exists in blood vessels within demyelinating lesions and suggests that SWI might provide a useful therapeutic tool to estimate the level of deoxyhemoglobin (and thus hypoxia) in MS patients. Additional studies using novel pO₂ sensors implanted in the cerebral cortex and cerebellar gray matter of EAE rats demonstrated a strong correlation between the degree of white matter autoimmune inflammation and degree of hypoxia (Johnson et al., 2016). Consistent with these findings, a recent study used frequency domain nearinfrared spectroscopy (fdNRS) to measure microvascular hemoglobin oxygen saturation (StO₂) in the cerebral cortex of MS patients. This revealed that compared to control subjects, StO₂ levels in MS patients were markedly decreased, and a significant relationship between StO₂ and clinical disability was demonstrated, strongly supporting the concept of hypoxia as a pathogenic contributor in MS progression (Yang and Dunn, 2015). Taken together, these combined data demonstrate that a true state of hypoxia exists in MS lesions, but also suggest that this may be substantially augmented by a state of virtual hypoxia as a result of ROS and NO-mediated mitochondrial dysfunction.

3. Potential causes of hypoxia in the developing MS lesion

If hypoxia is a potential trigger of MS, then it is important to

first understand more about the factors that could lead to this state of hypoxia (see table 1).

3.1 The watershed concept

Almost sixty years ago, in a study of MS lesions in periventricular regions, Brownell and Hughes (1962) noted that the majority of demyelinating lesions in the brain occur in watershed areas between the anterior, middle, and posterior cerebral arteries, areas that are located at the terminal endpoints of the arterial branches and where the blood supply is at its lowest level (Brownell and Hughes, 1962; Haider et al., 2016). Equally, in the spinal cord, several studies have shown that demyelinating lesions tend to occur in white matter tracts or in regions around the gray-white matter border zone, areas that have the poorest blood supply, i.e., the lowest density of blood vessels (Hassler, 1966; Turnbull et al., 1966; Tveten, 1976; Koyanagi et al., 1993a, b; Halder et al., 2018b). More recently, an MRI study noted that lesions are most often found in cerebral regions that have relatively poor perfusion (Holland et al., 2012). Other studies have reinforced this concept by showing that cerebral perfusion in the brain of MS patients is markedly reduced compared to normal controls (Law et al., 2004; Adhya et al., 2006; Varga et al., 2009; Papadaki et al., 2012). Taken together, these findings support the concept that cerebral hypoperfusion may predispose to the development of MS lesions, and that specific watershed areas in the CNS may be more susceptible because of the inherent limitations of their blood supply. Thus, under normal circumstances, blood supply would be adequate to meet metabolic demands; but, when the blood (or oxygen) supply is reduced, these vulnerable regions would be the first to manifest hypoxic distress. Thus, lack of adequate blood supply predisposes to the development of focal demyelinating lesions.

3.2 Cerebral hypoperfusion due to vascular dysfunction

Strong evidence supports the notion that cerebral blood flow (CBF) is reduced in MS patients. This was first demonstrated more than 30 years ago by a cluster of studies using a combination of single-photon emission computed tomography and positron emission tomography (PET) (Swank et al., 1983; Brooks et al., 1984; Lycke et al., 1993; Sun et al., 1998). More

recently, the development of improved imaging analysis using dynamic susceptibility contrast-enhanced perfusion magnetic resonance imaging (DSC-MRI) has enabled investigators to distinguish between demyelinating lesions, normal appearing white matter (NAWM), and grey matter. Importantly, this revealed that CBF is reduced both in NAWM and in deep grey matter in patients with all different types of MS, including relapsing-remitting (R-R), primary progressive, and clinically isolated syndrome (Law et al., 2004; Adhya et al., 2006; Varga et al., 2009; Papadaki et al., 2012). These observations support the concept that cerebral hypoperfusion may be a universal early event in the pathogenesis of all types of MS. A recent MRI study confirmed similar reductions in cervical blood flow in MS patients (El Sankari et al., 2013). This raises the fundamental question: if MS is triggered by reduced CBF, then what is the cause of this reduced blood flow? It seems unlikely to be a result of reduced energy demand due to axonal degeneration because reduced CBF is an early event, being detected well before the relatively late appearance of axonal degeneration (D'Haeseleer et al., 2015).

In light of recent studies highlighting the high degree of comorbidity between vascular disease and MS, and elevated levels of cardiovascular risk factors in MS patients, one possibility is that disruption of vascular function may be a cause (Christiansen et al., 2010; Karmon et al., 2012; Tettey et al., 2014; Marrie et al., 2015; Kappus et al., 2016). In one study of children and teenagers, the relative risk of developing MS was found to be much higher in patients with preexisting type 1 diabetes than a control group (Bechtold et al., 2014). Dyslipidemia is also implicated by the finding that elevation of both total cholesterol and low-density lipoprotein cholesterol are strongly associated with both the first clinical episode of MS as well as a worsening of clinical status (Giubilei et al., 2002; Weinstock-Guttman et al., 2011; Weinstock-Guttman et al., 2013). Other reports have shown that another cardiovascular risk factor, cigarette smoking, is strongly associated with increased risk of MS in a dose-dependent manner (Hernán et al., 2001; Hedström et al., 2013; O'Gorman et al., 2014). Importantly, MS patients show a higher incidence of ischemic stroke (Capkun et al., 2015; Tseng et al., 2015; Hong et al., 2019). Taken together, these combined data support the idea that vascular pathology may be an important pre-disposing factor in MS pathogenesis.

In an alternative explanation, it's interesting to compare MS with another neurological disease, migraine, which also shows a female:male incidence ratio of 3:1, as well as predominantly affecting the young-middle age population (Burch et al., 2018). Interestingly, several studies have pointed to the clinical overlap between migraine and MS (Elliott, 2007; Applebee, 2012), with the incidence of migraine 3-fold higher in MS patients, and the presence of migraine being a predictor of a more symptomatic MS course (Kister et al., 2010). Other studies have suggested that migraine with aura may be an early symptom of MS and/ or an indicator of exacerbation of MS symptoms (Tabby et al., 2013). In addition, migraineurs have an elevated risk of developing MS compared to the control population (Kister et al., 2012). In migraine pathophysiology, a long-held belief is that defective autoregulation of CBF may be responsible (Blau, 1978; Lauritzen, 1987) while other studies have described intimal thickening in the cerebral arterioles of migraine patients (Poyrazoglu et al., 2016; Wang et al., 2019). So, could the reduced CBF in MS patients be the result of abnormalities in structure or function of cerebral blood vessels? In support of this concept, studies have demonstrated intimal thickening of cerebral arteries (Yuksel et al., 2019) as well as stiffer retinal arterioles in MS patients (Kochkorov et al., 2009), while at the functional level, impaired vasodilatory responses in cerebral arterioles and reduced levels of reactive hyperemia have been

described in MS patients (Ranadive et al., 2012; Marshall et al., 2014).

At the molecular level, one mechanism that may link neuroinflammation and reduced blood flow is based on the finding that MS patients show impaired vasodilatory responses in cerebral arterioles (Marshall et al., 2014). Endothelin-1 (ET-1) is a vasoconstrictive peptide that is normally produced by endothelial cells but not glial cells (Piechota et al., 2010). Interestingly, ET-1 levels in peripheral blood and cerebrospinal fluid (CSF) are elevated in MS patients (Speciale et al., 2000; Haufschild et al., 2001), and ET-1 expression, normally absent in resting astrocytes, is upregulated in reactive astrocytes (Nie and Olsson, 1996; Ostrow et al., 2000). In support of this concept, a recent study showed that CBF in MS patients was reduced by approximately 20% in many brain regions and importantly, pharmacological blockade of the ET-1 receptor restored CBF to control levels (D'Haeseleer et al., 2013). Based on these findings, one mechanism that has been proposed is that early activation of astrocytes in MS lesions triggers ET-1 expression, which enhances arteriolar constriction, thus reducing CBF (D'Haeseleer et al., 2013).

3.3 Vascular inflammation

The idea that vascular dysfunction may be an early pivotal trigger of MS pathogenesis is further supported by the key observation that in acute demyelinating MS lesions, endothelial cell activation, as indicated by upregulation of MHC class II and fibrin deposition, is observed prior to parenchymal reaction or demyelination (Wakefield et al., 1994). Endothelial activation leads to upregulation of adhesion molecules such as selectins, vascular cell adhesion molecule (VCAM)-1, and intercellular adhesion molecule (ICAM)-1, which promote T lymphocyte adherence to the luminal side of endothelial cells (Engelhardt et al., 1994; Lee and Benveniste, 1999). This in turn, results in aggregation of lymphocyte clusters within the lumen of microvessels, that secrete a spectrum of proinflammatory cytokines and chemokines that further promote leukocyte extravasation and amplify the inflammatory cascade. Infiltrated leukocytes also secrete proteases, such as matrix metalloproteinase-9 (MMP-9), which can strip endothelial cells of critical cell surface proteins necessary for maintaining tight barrier integrity, e.g., tight junction proteins and adherens proteins (Redford et al., 1997b; Milner et al., 2007; Boroujerdi et al., 2015). In addition, the inflammatory environment can also promote release of vasoactive peptides, which impact vascular function by inhibiting homeostatic vasodilatory responses, leading to constriction of blood vessels and reduced blood flow. A good example is the vasoconstrictive peptide endothelin-1 (ET-1), which is strongly upregulated by reactive astrocytes, and whose vasoconstrictive action reduces CBF (D'Haeseleer et al., 2013; D'Haeseleer et al., 2015). Blood vessels within inflammatory regions can also be targeted for damage and destruction by several different components of the immune system (cytokines, antibodies or CD8+ T lymphocytes), or by virtue of their heightened activation state, inflamed blood vessels can trigger local activation of the clotting cascade resulting in thrombotic occlusion and ischemia (Tomasson et al., 2009; Emmi et al., 2015). This is because inflammation promotes a pro-thrombotic state as a result of downregulation of anti-thrombotic proteins such as thrombomodulin and upregulation of thrombotic factors, such as tissue factor and thrombin (Kopp et al., 1997; Esmon, 2005, 2012; Foley and Conway, 2016).

Furthermore, at the earliest stage of demyelinating lesion development, the BBB is disrupted (McDonald and Barnes, 1989; Kermode et al., 1990; Gay and Esiri, 1991; Colosimo et al., 1992; Kirk et al., 2003) resulting in extravascular leakage of serum proteins into the CNS, which perturbs tissue fluid homeostasis in the local environment. The ensuing localized edema compresses cerebral microvessels, resulting in ischemia and hypoxic tissue damage. In the clinical setting, BBB disruption is evaluated using gadolinium-enhanced MRI, which provides one of the earliest markers of impending tissue damage and is currently the best prognostic tool for monitoring the appearance of new active MS lesions (McDonald and Barnes, 1989; Colosimo et al., 1992; McDonald, 1994; Compston and Coles, 2008).

3.4 Systemic hypoxia

In addition to being the result of reduced blood flow, tissue hypoxia can also occur when blood flow is relatively normal, but the body is in a state of systemic hypoxia. Many different medical conditions can lead to a hypoxic state, including obstructive sleep apnea, several types of lung disease, including acute (bronchitis, pneumonia) and chronic (asthma, pulmonary fibrosis, emphysema) as well as reduced cardiac function (Bandopadhyay and Selvamurthy, 2000; Yaffe et al., 2011; Bucks et al., 2013; Leng et al., 2017; Yohannes et al., 2017; Russ et al., 2020). When systemic hypoxia occurs, areas of the CNS with the poorest blood supply (watershed areas) will be most vulnerable and will therefore be the first to display signs of hypoxia. Interestingly, epidemiological studies have highlighted a strong association between sleep apnea and MS, in that sleep apnea is much more common in MS patients compared to control subjects (Braley et al., 2012), prompting the question: what accounts for the increased incidence of sleep apnea in MS patients? However, considering the emerging evidence implicating hypoxia as a potential trigger of MS, it might be equally insightful to ask the reverse question.

4. The protective effect of hypoxic conditioning in animal models of demyelinating disease

Inspired by overwhelming evidence demonstrating the strong neuroprotective influence of hypoxic pre-conditioning in animal models of ischemic stroke (Dowden and Corbett, 1999; Miller et al., 2001; Stowe et al., 2011), the Dore-Duffy lab was the first to examine whether hypoxic pre-conditioning might also reduce the clinical severity of EAE. Using a chronic progressive EAE model (MOG35-55 peptide immunogen in C57BL6/J mice), they demonstrated that mice maintained under chronic mild hypoxic conditions $(10\% O_2)$ from the time of immunization (i.e., before EAE was manifest), showed reduced levels of neurological disability, which correlated with reduced levels of leukocyte infiltration into the spinal cord (Dore-Duffy et al., 2011). In a subsequent study, the same group found that chronic mild hypoxia (CMH) reduced the degree of CD4⁺ T lymphocyte infiltration into the spinal cord and this correlated with a delayed Th17-specific cytokine response. Conversely, spinal cords of CMH-treated mice contained increased numbers of CD4⁺/CD25⁺/FoxP3⁺ regulatory T cells (Tregs) as well as higher levels of the anti-inflammatory cytokine IL-10 (Esen et al., 2016). Based on these findings, the authors concluded that CMH triggers a number of endogenous adaptations that promotes the establishment of an anti-inflammatory milieu within the CNS.

In a previous series of studies, we demonstrated that CMH $(8-10\% O_2)$ promotes favorable changes in cerebrovascular properties that includes a marked angiogenic remodeling response, resulting in enhanced vessel density as well as marked upregulation of endothelial tight junction proteins, suggestive of increased vascular integrity (Milner et al., 2008; Li et al., 2010; Boroujerdi and Milner, 2015; Halder et al., 2018b). Based on these findings, we hypothesized that CMH might confer protection from EAE, in part via its positive influence on vascular integrity. In contrast to the Dore-Duffy lab, we chose to perform our hypoxic conditioning studies in

the relapsing-emitting (R-R) EAE model (PLP139-151 peptide immunogen in SJL/J mice) for two reasons; first, this model is more representative of the most common form of MS seen in patients (85% get the R-R form), and second, this model facilitates analysis of an intervention not just on peak disease but also on the extent of remission, and on rates of relapse. Strikingly, we found that when started at the same time as EAE induction, CMH significantly reduced the severity of peak disease (by more than 50%), with protection maintained for the duration of the experiment (7 weeks) (Halder et al., 2018a). This was matched at the histological level by reduced levels of BBB breakdown, leukocyte infiltration, and demyelination. Tissue analysis revealed that hypoxia-enhanced BBB stability correlated with reduced expression of the endothelial activation leukocyte-docking molecules, vascular cell adhesion molecule (VCAM)-1 and intercellular adhesion molecule (ICAM)-1, as well as increased expression of the endothelial tight junction proteins occludin and ZO-1. CMH also triggered increased levels of the extracellular matrix (ECM) protein laminin-111 in the vascular basement membrane, which has been shown to inhibit leukocyte transmigration (Sixt et al., 2001). Based on these findings, we concluded that the protective effect of hypoxic pre-conditioning in EAE is underpinned, at last in part, by several different changes in blood vessel properties that contribute to enhanced vascular integrity (Halder et al., 2018a).

More recently, we investigated the clinically important question of whether CMH can impact the progression of preexisting EAE. Using the R-R EAE model, we found that when mice with significant neurological disability were treated with CMH, while this had no impact on the peak disease score, CMH markedly accelerated clinical recovery during the remission phase of disease, resulting in long-term stable reductions in neurological deficit (Halder and Milner, 2020). At the histological level, this correlated with marked reductions in vascular disruption, leukocyte accumulation and demyelination within the spinal cord. Mechanistically, we found that CMH reduced endothelial expression of VCAM-1 while promoting faster re-expression of the tight junction proteins occludin and ZO-1 following their disappearance during the peak phase of disease. Most interestingly, while CMH did not influence the level of peak clinical disease or the degree of inflammatory leukocytosis at peak disease (3 days after CMH started), apoptotic removal of infiltrated leukocytes during the remission phase was profoundly enhanced, correlating with marked clinical improvement. Within the EAE spinal cord, HIF-1 α was strongly expressed in infiltrated monocytes and the number of HIF-1 α -positive monocytes was greatly increased by CMH. Taken together, these findings are consistent with our previous data in suggesting that CMH confers protection from EAE, in part by enhancing different mechanisms contributing to vascular integrity. They also make the important point that CMH greatly accelerates monocyte apoptotic removal within acute inflammatory lesions by amplifying the hypoxic stress manifest within these cells.

5. Which molecular mechanisms account for the protective effect of hypoxic conditioning?

The studies described to date have identified three potential mechanisms that may underlie the protective effect of hypoxic conditioning on inflammatory demyelinating disease: (i) modulation of the peripheral immune system, (ii) enhanced BBB integrity, and (iii) accelerated apoptotic removal of infiltrated monocytes (see Figure 1).

5.1 The influence of hypoxia on the immune system

In their first study, documenting the protective effect of CMH on the progression of EAE, the Dore-Duffy lab correlated



Figure 1. Summary of the proposed molecular mechanisms underlying the protective influence of hypoxic conditioning in MS. These include: (1) promoting vascular remodeling, which will accelerate vascular repair following leukocyte-induced breakdown as well as increase removal of waste products via increased vascularity, (2) enhanced vascular integrity due to upregulation of tight junction proteins and vascular basal lamina ECM protein expression (e.g., laminin), (3) promoting an anti-inflammatory environment within the CNS by shifting the balance between pathogenic Th17 encephalitogenic T cells and immunosuppressive regulatory T cells, and (4) promoting apoptosis of infiltrated monocytes.

reduced clinical disease with reduced leukocyte infiltration into the spinal cord (Dore-Duffy et al., 2011). In their second study, they correlated this protection with reduced numbers of CD4+ lymphocyte infiltration into the spinal cord, which was associated with a decreased destructive Th17-specific response, but interestingly, they also noted increased numbers of immunosuppressive regulatory T cells (Tregs) and increased levels of the anti-inflammatory cytokine interleukin (IL)-10 within the spinal cord (Esen et al., 2016). Based on these observations, the authors concluded that by tilting the Th17/Treg balance in favor of Tregs, CMH triggers an anti-inflammatory environment within the CNS. When considering how this might be achieved, three possible explanations spring to mind; first, that CMH exerts an anti-inflammatory effect on the whole immune system and what occurs in the spinal cord is simply a reflection of what is happening elsewhere in the body, second, that CMH alters the ability of the BBB to selectively regulate which types of immune cell enter the CNS, and third, that the balance between Th17 and Treg cells is regulated at the local level within the CNS

The influence of hypoxia on cells of the immune system is complex and depends on several different factors, including the immune cell type, severity and duration of hypoxia, as well as the inflammatory context. Broadly speaking, hypoxia stimulates the function of many innate immune cell types such as monocyte and neutrophils, as seen by increased chemotaxis and phagocytosis, and enhanced production of ROS and proinflammatory cytokines such as IL-6, which all serve to promote clearance of pathogens (Scannell et al., 1993; Klausen et al., 1997; Hartmann et al., 2000; Wang and Liu, 2009; Kiers et al., 2016). However, the impact of hypoxia on lymphocyte function is less clear. One study demonstrated that hypoxia drives T cell differentiation towards the Th17 effector pathway and away from Treg differentiation, an effect that was substantiated by the finding that transgenic mice with HIF-1-deficient T cells are resistant to EAE induction due to defective generation of Th17 effector cells (Dang et al., 2011). On the other hand, another group studying these events in intestinal mucosa showed that hypoxia tilts the balance in favor of Tregs, and that HIF-1a deletion resulted in reduced numbers of Tregs and a failure to control T cell-mediated inflammatory colitis (Clambey et al., 2012). Interestingly, Esen et al. (2016) showed that in the EAE model, systemic hypoxia had no impact on MOG-specific T cell sensitization or proliferation, suggesting that T cell antigen-specific functions per se are not altered by hypoxia. Clearly, more studies are required to clarify how hypoxia modulates the Th17/Treg balance during EAE and to understand whether this is a direct effect on T cells or an indirect effect via the influence of hypoxia on other CNS-resident cell types.

5.2 The hypoxic influence on vascular remodeling and vascular integrity

It is well established that CMH induces a robust vascular remodeling response in the CNS, resulting in enhanced vascularity as well as upregulation of several components of the BBB (Milner et al., 2008; Li et al., 2010; Boroujerdi and Milner, 2015; Halder et al., 2018b). Interestingly, in our recent study demonstrating the therapeutic benefit of hypoxic treatment of pre-existing EAE, we showed that the hypoxiainduced angiogenic response proceeds, even in the middle of a full-blown inflammatory response seen in EAE (Halder and Milner, 2020). Surprisingly, this vascular remodeling did not lead to more BBB disruption as might be predicted when endothelial cells uncouple from their neighbors during the angiogenic process, but instead resulted in enhanced vascular integrity. So how can vascular remodeling be beneficial in EAE? We think three potential reasons might account for this. First, one possibility is that when EAE occurs on a proangiogenic background, because the angiogenic process is already up and running, any blood vessels disrupted because of leukocyte infiltration, will be more quickly repaired and sealed. This idea is supported by our recent finding that transgenic mice with endothelial deletion of the angiogenic integrin $\alpha 5\beta 1$ (α 5-EC-KO mice), which show a delayed angiogenic response to CMH (Li et al., 2012), manifest earlier clinical disease along with greater BBB disruption in the EAE model (Kant et al., 2019).

Second, it's plausible that the increased vessel density improves the perfusion of affected tissue as well as speeding the removal of any waste/myelin breakdown products or inflammatory cytokines that are facilitating the inflammatory cycle. This idea is consistent with previous studies in MS patients, which demonstrated that enhanced vasodilation and increased blood flow triggered by factors such as histamine and amyl nitrate provided short-term clinical relief of symptoms, described as "relief by flush" (Jonez, 1948; Brickner, 1955; Brickner, 1958). It would also be in keeping with the "clean up" hypothesis, whereby it is suggested that angiogenesis is promoted under inflammatory conditions in part, as an attempt to remove tissue debris (Manoonkitiwongsa et al., 2001).

The third possibility is that CMH promotes BBB integrity. In a series of studies in non-EAE conditions, we have shown that CMH enhances both endothelial expression of tight junction proteins on CNS blood vessels, as well as increasing expression of the leukocyte-inhibitory ECM protein laminin within the vascular basal lamina (Li et al., 2010; Boroujerdi and Milner, 2015; Halder et al., 2018b; Halder et al., 2018c). Our recent work demonstrates that CMH triggers the same mechanisms during EAE progression (Halder et al., 2018a; Halder and Milner, 2020). In addition, CMH also reduces expression of the endothelial activation molecules VCAM-1 and ICAM-1 (Halder et al., 2018a; Halder and Milner, 2020), suggesting that CMH acts to dampen the vascular inflammatory response induced by EAE, while simultaneously reducing leukocyte extravasation across the BBB. Indeed, in our recent study describing the protective influence of CMH on pre-existing EAE, the impact on suppressing endothelial VCAM-1 expression was the earliest detectable event in CMH-treated mice (Halder and Milner, 2020). These findings are consistent with the marked impact of CMH on reducing vascular disruption and preventing leukocyte infiltration into the CNS.

5.3 Hypoxia accelerates the apoptosis of infiltrated leukocytes

One of the most notable findings from our recent EAE study was that hypoxic treatment of pre-existing disease strongly enhanced apoptotic removal of infiltrated monocytes from the spinal cord during the remission phase (Halder and Milner, 2020). Interestingly, in comparison to CD4⁺ T cells, which were evenly distributed throughout the entire spinal cord, monocytes accumulated in high-density cellular aggregates within the white matter and displayed the highest levels of HIF- 1α expression and apoptotic cell death. This is consistent with the idea that after entering the CNS parenchyma, infiltrated monocytes continue to proliferate and transform into activated, highly metabolic cells, to create high-density aggregates where hypoxia is most severe (Le Moan et al., 2015; Halder and Milner, 2020). Based on this, it is logical to assume that CMH will greatly enhance the hypoxia already experienced by these monocytes, and as suggested by previous studies, will trigger apoptotic pathways (Biju et al., 2004; Krick et al., 2005; Luo et al., 2006). This concept is supported by the observation that inhibition of caspase-mediated apoptosis prevents clinical remission in EAE (Okuda et al., 2000). Taken together, this data suggests that approaches designed to enhance monocyte hypoxia or apoptosis by other means (e.g., activating caspase-3 activation) could offer new ways forward in promoting clinical recovery in MS.

6. Does hypoxic conditioning hold therapeutic potential in MS?

As animal EAE studies have demonstrated the clinical efficacy of hypoxic conditioning, not just in pre-conditioning, but also more recently in the treatment of pre-existing disease (Esen et al., 2016; Halder et al., 2018a; Halder and Milner, 2020), how might this be exploited for therapeutic potential? Two possible approaches are worth considering. First, a growing number of reports in other neurological diseases including ischemic stroke, Alzheimer's disease, spinal cord injury and epilepsy, have demonstrated the therapeutic potential of intermittent hypoxic training (IHT), which consists of regular, short hypoxic exposures (Fuller et al., 2003; Golder and Mitchell, 2005; Manukhina et al., 2010; Stowe et al., 2011; Zhen et al., 2014). Because CMH promotes long term adaptive changes in vascular (enhanced vascularity and vascular integrity) (Milner et al., 2008; Li et al., 2010; Boroujerdi and Milner, 2015; Halder et al., 2018b) and immunological properties (Esen et al., 2016), it's possible that these changes may provide benefit in treating MS. Before this is ready to test in MS patients however, it will first be necessary to demonstrate proof of the IHT concept in EAE studies, something that has not yet been examined, as well as define an optimal IHT protocol that would provide maximum benefit. Second, rather than using hypoxia itself, drugs called hypoxia mimetics could be used to stimulate hypoxic signaling pathways. These drugs inhibit the prolyl hydroxylase domain (PHD) group of enzymes, which prevents hydroxylation of HIF- 1α , thus raising HIF-1 α levels (Jain et al., 2016). Interestingly, recent studies have demonstrated that the cannabinoid VCE-004.8, which has hypoxic mimetic activity, reduces the severity of EAE (Navarrete et al., 2018), suggesting promising clinical potential for this therapeutic approach.

7. Conclusions

Recent studies have demonstrated that hypoxic conditioning is effective in reducing clinical severity in the EAE animal model of MS, not just as pre-conditioning, but more importantly, when used to treat pre-existing disease. Mechanistic studies have suggested that this protection is mediated by beneficial adaptive changes in the properties of blood vessels and the immune system. More studies are now needed to further clarify these changes and define the key intracellular signaling pathways responsible. At the same time, current data suggests it may be fruitful to explore the therapeutic potential of this phenomenon by examining the impact of intermittent hypoxic training (IHT) and hypoxia mimetics in EAE.

References

- Aboul-Enein F, Lassmann H (2005) Mitochondrial damage and histotoxic hypoxia: a pathway of tissue injury in inflammatory brain disease? Acta Neuropathol 109:49-55.
- Aboul-Enein F, Rauschka H, Kornek B, Stadelmann C, Stefferl A, Brück W, Lucchinetti C, Schmidbauer M, Jellinger K, Lassmann H (2003) Preferential loss of myelin-associated glycoprotein reflects hypoxia-like white matter damage in stroke and inflammatory brain diseases. J Neuropathol Exp Neurol 62:25-33.
- Adhya S, Johnson G, Herbert J, Jaggi H, Babb JS, Grossman RI, Inglese M (2006) Pattern of hemodynamic impairment in multiple sclerosis: dynamic susceptibility contrast perfusion MR imaging at 3.0 T. Neuroimage 33:1029-1035.
- Applebee A (2012) The clinical overlap of multiple sclerosis and headache. Headache 52 Suppl 2:111-116.
- Babbe H, Roers A, Waisman A, Lassmann H, Goebels N, Hohlfeld R, Friese M, Schröder R, Deckert M, Schmidt S, Ravid R, Rajewsky K (2000) Clonal expansions of CD8(+) T cells dominate the T cell infiltrate in active multiple sclerosis lesions as shown by micromanipulation and single cell polymerase chain reaction. J Exp Med 192:393-404.
- Bandopadhyay P, Selvamurthy W (2000) Suggested predictive indices for high altitude pulmonary oedema. J Assoc Physicians India 48:290-294.
- Barnett MH, Prineas JW (2004) Relapsing and remitting multiple sclerosis: pathology of the newly forming lesion. Ann Neurol 55:458-468.

- Barnett MH, Parratt JD, Pollard JD, Prineas JW (2009) MS: is it one disease? Int MS J 16:57-65.
- Bechtold S, Blaschek A, Raile K, Dost A, Freiberg C, Askenas M, Fröhlich-Reiterer E, Molz E, Holl RW (2014) Higher relative risk for multiple sclerosis in a pediatric and adolescent diabetic population: analysis from DPV database. Diabetes Care 37:96-101.
- Biju MP, Neumann AK, Bensinger SJ, Johnson RS, Turka LA, Haase VH (2004) Vhlh gene deletion induces Hif-1-mediated cell death in thymocytes. Mol Cell Biol 24:9038-9047.
- Blau JN (1978) Migraine: A vasomotor instability of the meningeal circulation. Lancet 2:1136-1139.
- Boroujerdi A, Milner R (2015) Defining the critical hypoxic threshold that promotes vascular remodeling in the brain. Exp Neurol 263:132-140.
- Boroujerdi A, Welser-Alves J, Milner R (2015) Matrix metalloproteinase-9 mediates post-hypoxic vascular pruning of cerebral blood vessels by degrading laminin and claudin-5. Angiogenesis Mar 27 Epub.
- Braley TJ, Segal BM, Chervin RD (2012) Sleep-disordered breathing in multiple sclerosis. Neurology 79:929-936.
- Brickner RM (1955) Phenomenon of relief by flush in multiple sclerosis; its use as a foundation for therapy. AMA Arch Neurol Psychiatry 73:232-240.
- Brickner RM (1958) Pharmacological reduction of abnormality in multiple sclerosis within minutes: a statistical study. J Nerv Ment Dis 127:308-322.
- Brooks DJ, Leenders KL, Head G, Marshall J, Legg NJ, Jones T (1984) Studies on regional cerebral oxygen utilisation and cognitive function in multiple sclerosis. J Neurol Neurosurg Psychiatry 47:1182-1191.
- Brownell B, Hughes JT (1962) The distribution of plaques in the cerebrum in multiple sclerosis. J Neurol Neurosurg Psychiatry 25:315-320.
- Brownlee WJ, Hardy TA, Fazekas F, Miller DH (2017) Diagnosis of multiple sclerosis: progress and challenges. Lancet 389:1336-1346.
- Brück W, Porada P, Poser S, Rieckmann P, Hanefeld F, Kretzschmar HA, Lassmann H (1995) Monocyte/ macrophage differentiation in early multiple sclerosis lesions. Ann Neurol 38:788-796.
- Bucks RS, Olaithe M, Eastwood P (2013) Neurocognitive function in obstructive sleep apnoea: a meta-review. Respirology 18:61-70.
- Burch R, Rizzoli P, Loder E (2018) The Prevalence and Impact of Migraine and Severe Headache in the United States: Figures and Trends From Government Health Studies. Headache 58:496-505.
- Capkun G, Dahlke F, Lahoz R, Nordstrom B, Tilson HH, Cutter G, Bischof D, Moore A, Simeone J, Fraeman K, Bancken F, Geissbühler Y, Wagner M, Cohan S (2015) Mortality and comorbidities in patients with multiple sclerosis compared with a population without multiple sclerosis: An observational study using the US Department of Defense administrative claims database. Mult Scler Relat Disord 4:546-554.
- Christiansen CF, Christensen S, Farkas DK, Miret M, Sørensen HT, Pedersen L (2010) Risk of arterial cardiovascular diseases in patients with multiple sclerosis: a population-based cohort study. Neuroepidemiology 35:267-274.
- Clambey ET, McNamee EN, Westrich JA, Glover LE, Campbell EL, Jedlicka P, de Zoeten EF, Cambier JC, Stenmark KR, Colgan SP, Eltzschig HK (2012) Hypoxia-

inducible factor-1 alpha-dependent induction of FoxP3 drives regulatory T-cell abundance and function during inflammatory hypoxia of the mucosa. Proc Natl Acad Sci U S A 109:E2784-2793.

- Colosimo M, Amatruda A, Cioffi RP (1992) Magnetic resonance imaging in multiple sclerosis: an overview. Ital J Neurol Sci 13:113-123.
- Compston A, Coles A (2008) Multiple sclerosis. Lancet 372:1502-1517.
- D'Haeseleer M, Hostenbach S, Peeters I, Sankari SE, Nagels G, De Keyser J, D'Hooghe M B (2015) Cerebral hypoperfusion: a new pathophysiologic concept in multiple sclerosis? J Cereb Blood Flow Metab 35:1406-1410.
- D'Haeseleer M, Beelen R, Fierens Y, Cambron M, Vanbinst AM, Verborgh C, Demey J, De Keyser J (2013) Cerebral hypoperfusion in multiple sclerosis is reversible and mediated by endothelin-1. Proc Natl Acad Sci U S A 110:5654-5658.
- Dang EV, Barbi J, Yang HY, Jinasena D, Yu H, Zheng Y, Bordman Z, Fu J, Kim Y, Yen HR, Luo W, Zeller K, Shimoda L, Topalian SL, Semenza GL, Dang CV, Pardoll DM, Pan F (2011) Control of T(H)17/T(reg) balance by hypoxia-inducible factor 1. Cell 146:772-784.
- Davies AL, Desai RA, Bloomfield PS, McIntosh PR, Chapple KJ, Linington C, Fairless R, Diem R, Kasti M, Murphy MP, Smith KJ (2013) Neurological deficits caused by tissue hypoxia in neuroinflammatory disease. Ann Neurol 74:815-825.
- Desai RA, Davies AL, Tachrount M, Kasti M, Laulund F, Golay X, Smith KJ (2016) Cause and prevention of demyelination in a model multiple sclerosis lesion. Ann Neurol 79:591-604.
- Diaz-Villoslada P, Shih A, Shao L, Genain CP, Hauser SL (1999) Autoreactivity to myelin antigens: myelin/oligodendrocyte glycoprotein is a prevalent autoantigen. J Neuroimmunol 99:36-43.
- Dore-Duffy P, Wencel M, Katyshev V, Cleary K (2011) Chronic mild hypoxia ameliorates chronic inflammatory activity in myelin oligodendrocyte glycoprotein (MOG) peptide induced experimental autoimmune encephalomyelitis (EAE). Adv Exp Med Biol 701:165-173.
- Doshi A, Chataway J (2017) Multiple sclerosis, a treatable disease. Clin Med (Lond) 17:530-536.
- Dowden J, Corbett D (1999) Ischemic preconditioning in 18- to 20-month-old gerbils: long-term survival with functional outcome measures. Stroke 30:1240-1246.
- Dutta R, McDonough J, Yin X, Peterson J, Chang A, Torres T, Gudz T, Macklin WB, Lewis DA, Fox RJ, Rudick R, Mirnics K, Trapp BD (2006) Mitochondrial dysfunction as a cause of axonal degeneration in multiple sclerosis patients. Ann Neurol 59:478-489.
- El Sankari S, Balédent O, van Pesch V, Sindic C, de Broqueville Q, Duprez T (2013) Concomitant analysis of arterial, venous, and CSF flows using phase-contrast MRI: a quantitative comparison between MS patients and healthy controls. J Cereb Blood Flow Metab 33:1314-1321.
- Elliott DG (2007) Migraine in multiple sclerosis. Int Rev Neurobiol 79:281-302.
- Emmi G, Silvestri E, Squatrito D, Amedei A, Niccolai E, D'Elios MM, Della Bella C, Grassi A, Becatti M, Fiorillo C, Emmi L, Vaglio A, Prisco D (2015) Thrombosis in vasculitis: from pathogenesis to treatment. Thromb J 13:15.

Engelhardt B, Conley FK, Butcher EC (1994) Cell adhesion

molecules on vessels during neuroinflammation in the mouse central nervous system. J Neuroimmunol 51:199-208.

- Esen N, Katyshev V, Serkin Z, Katysheva S, Dore-Duffy P (2016) Endogenous adaptation to low oxygen modulates T-cell regulatory pathways in EAE. J Neuroinflammation 13:13.
- Esmon CT (2005) Coagulation inhibitors in inflammation. Biochem Soc Trans 33:401-405.
- Esmon CT (2012) Protein C anticoagulant system--antiinflammatory effects. Semin Immunopathol 34:127-132.
- French-Constant C (1994) Pathogenesis of multiple sclerosis. Lancet 343:271-275.
- Foley JH, Conway EM (2016) Cross Talk Pathways Between Coagulation and Inflammation. Circ Res 118:1392-1408.
- Fuller DD, Johnson SM, Olson EBJ, Mitchell GS (2003) Synaptic pathways to phrenic motorneurons are enhanced by chronic intermittent hypoxia after cervical spinal cord injury. J Neurosci 23:2993-3000.
- Gay D, Esiri M (1991) Blood-Brain Barrier Damage in Acute Multiple Sclerosis Plaques. Brain 114:557-572.
- Gay FW, Drye TJ, Dick GW, Esiri MM (1997) The application of multifactorial cluster analysis in the staging of plaques in early multiple sclerosis. Identification and characterization of the primary demyelinating lesion. Brain 120 (Pt 8):1461-1483.
- Genain CP, Cannella B, Hauser SL, Raine CS (1999) Identification of autoantibodies associated with myelin damage in multiple sclerosis. Nat Med 5:170-175.
- Giubilei F, Antonini G, Di Legge S, Sormani MP, Pantano P, Antonini R, Sepe-Monti M, Caramia F, Pozzilli C (2002) Blood cholesterol and MRI activity in first clinical episode suggestive of multiple sclerosis. Acta Neurol Scand 106:109-112.
- Golder FJ, Mitchell GS (2005) Spinal synaptic enhancement with acute intermittent hypoxia imrpoves respiratory function after chronic cervical spinal cord injury. J Neurosci 25:2925-2932.
- Graumann U, Reynolds R, Steck AJ, Schaeren-Wiemers N (2003) Molecular changes in normal appearing white matter in multiple sclerosis are characteristic of neuroprotective mechanisms against hypoxic insult. Brain Pathol 13:554-573.
- Haider L, Zrzavy T, Hametner S, Höftberger R, Bagnato F, Grabner G, Trattnig S, Pfeifenbring S, Brück W, Lassmann H (2016) The topograpy of demyelination and neurodegeneration in the multiple sclerosis brain. Brain 139:807-815.
- Halder SK, Milner R (2020) Chronic mild hypoxia accelerates recovery from pre-existing EAE by enhancing vascular integrity and apoptosis of infiltrated monocytes. Proc Natl Acad Sci U S A 117:11126-11135.
- Halder SK, Kant R, Milner R (2018a) Hypoxic pre-conditioning suppresses experimental autoimmune encephalomyelitis by modifying multiple properties of blood vessels. Acta Neuropathologica Communications 6:86.
- Halder SK, Kant R, Milner R (2018b) Chronic mild hypoxia promotes profound vascular remodeling in spinal cord blood vessels, preferentially in white matter, via an α 5 β 1 integrin-mediated mechanism. Angiogenesis 21:251-266.
- Halder SK, Kant R, Milner R (2018c) Chronic mild hypoxia increases expression of laminins 111 and 411 and the laminin receptor $\alpha 6\beta 1$ integrin at the blood-brain barrier. Brain Res 1700:78-85.

- Hartmann G, Tschöp M, Fischer R, Bidlingmaier C, Riepl R, Tschöp K, Hautmann H, Endres S, Toepfer M (2000) High altitude increases circulating interleukin-6, interleukin-1 receptor antagonist and C-reactive protein. Cytokine 12:246-252.
- Hassler O (1966) Blood supply to human spinal cord. A microangiographic study. Arch Neurol 15:302-307.
- Haufschild T, Shaw SG, Kesselring J, Flammer J (2001) Increased endothelin-1 plasma levels in patients with multiple sclerosis. J Neuroophthalmol 21:37-38.
- Hedström AK, Hillert J, Olsson T, Alfredsson L (2013) Smoking and multiple sclerosis susceptibility. Eur J Epidemiol 28:867-874.
- Hernán MA, Olek MJ, Ascherio A (2001) Cigarette smoking and incidence of multiple sclerosis. Am J Epidemiol 154:69-74.
- Holland CM, Charil A, Csapo I, Liptak Z, Ichise M, Khoury SJ, Bakshi R, Weiner HL, Guttmann CR (2012) The relationship between normal cerebral perfusion patterns and white matter lesion distribution in 1,249 patients with multiple sclerosis. J Neuroimaging 22:129-136.
- Hong Y, Tang HR, Ma M, Chen N, Xie X, He L (2019) Multiple sclerosis and stroke: a systematic review and metaanalysis. BMC Neurol 19:139.
- Itoyama Y, Sternberger NH, Webster HD, Quarles RH, Cohen SR, Richardson EP, Jr. (1980) Immunocytochemical observations on the distribution of myelin-associated glycoprotein and myelin basic protein in multiple sclerosis lesions. Ann Neurol 7:167-177.
- Itoyama Y, Webster HD, Sternberger NH, Richardson EP, Jr., Walker DL, Quarles RH, Padgett BL (1982) Distribution of papovavirus, myelin-associated glycoprotein, and myelin basic protein in progressive multifocal leukoencephalopathy lesions. Ann Neurol 11:396-407.
- Jain IH, Zazzeron L, Goli R, Alexa K, Schatzman-Bone S, Dhillon H, Goldberger O, Peng J, Shalem O, Sanjana NE, Zhang F, Goessling W, Zapol WM, Mootha VK (2016) Hypoxia as a therapy for mitochondrial disease. Science 352:54-61.
- Johnson TW, Wu Y, Nathoo N, Rogers JA, Wee Yong V, Dunn JF (2016) Gray Matter Hypoxia in the Brain of the Experimental Autoimmune Encephalomyelitis Model of Multiple Sclerosis. PLoS One 11:e0167196.
- Jonez HD (1948) Multiple sclerosis; treatment with histamine and d-tubocurarine. Ann Allergy 6:550-563.
- Kant R, Halder SK, Bix GJ, Milner R (2019) Absence of endothelial α 5 β 1 integrin triggers early onset of experimental autoimmune encephalomyelitis due to reduced vascular remodeling and compromised vascular integrity. Acta Neuropathologica Communications 7:11.
- Kappus N, Weinstock-Guttman B, Hagemeier J, Kennedy C, Melia R, Carl E, Ramasamy DP, Cherneva M, Durfee J, Bergsland N, Dwyer MG, Kolb C, Hojnacki D, Ramanathan M, Zivadinov R (2016) Cardiovascular risk factors are associated with increased lesion burden and brain atrophy in multiple sclerosis. J Neurol Neurosurg Psychiatry 87:181-187.
- Karmon Y, Ramanathan M, Minagar A, Zivadinov R, Weinstock-Guttman B (2012) Arterial, venous and other vascular risk factors in multiple sclerosis. Neurol Res 34:754-760.
- Kermode AG, Thompson AJ, Tofts P, MacManus DG, Kendall BE, Kingsley DP, Moseley IF, Rudge P, McDonald WI (1990) Breakdown of the blood-brain barrier precedes

symptoms and other MRI signs of new lesions in multiple sclerosis. Pathogenetic and clinical implications. Brain 113 (Pt 5):1477-1489.

- Kiers HD, Scheffer GJ, van der Hoeven JG, Eltzschig HK, Pickkers P, Kox M (2016) Immunologic Consequences of Hypoxia during Critical Illness. Anesthesiology 125:237-249.
- Kirk J, Plumb J, Mirakhur M, McQuaid S (2003) Tight junction abnormality in multiple sclerosis white matter affects all calibres of vessel and is associated with bloodbrain barrier leakage and active demyelination. J Pathol 201:319-327.
- Kister I, Munger KL, Herbert J, Ascherio A (2012) Increased risk of multiple sclerosis among women with migraine in the Nurses' Health Study II. Mult Scler 18:90-97.
- Kister I, Caminero AB, Monteith TS, Soliman A, Bacon TE, Bacon JH, Kalina JT, Inglese M, Herbert J, Lipton RB (2010) Migraine is comorbid with multiple sclerosis and associated with a more symptomatic MS course. J Headache Pain 11:417-425.
- Klausen T, Olsen NV, Poulsen TD, Richalet JP, Pedersen BK (1997) Hypoxemia increases serum interleukin-6 in humans. Eur J Appl Physiol Occup Physiol 76:480-482.
- Kochkorov A, Gugleta K, Kavroulaki D, Katamay R, Weier K, Mehling M, Kappos L, Flammer J, Orgül S (2009) Rigidity of retinal vessels in patients with multiple sclerosis. Klin Monbl Augenheilkd 226:276-279.
- König FB, Wildemann B, Nessler S, Zhou D, Hemmer B, Metz I, Hartung HP, Kieseier BC, Brück W (2008) Persistence of immunopathological and radiological traits in multiple sclerosis. Arch Neurol 65:1527-1532.
- Kopp CW, Siegel JB, Hancock WW, Anrather J, Winkler H, Geczy CL, Kaczmarek E, Bach FH, Robson SC (1997) Effect of porcine endothelial tissue factor pathway inhibitor on human coagulation factors. Transplantation 63:749-758.
- Koyanagi I, Tator CH, Lea PJ (1993a) Three-dimensional analysis of the vascular system in the rat spinal cord with scanning electron microscopy of vascular corrosion casts. Part 2: Acute spinal cord injury. Neurosurgery 33:285-291; discussion 292.
- Koyanagi I, Tator CH, Lea PJ (1993b) Three-dimensional analysis of the vascular system in the rat spinal cord with scanning electron microscopy of vascular corrosion casts. Part 1: Normal spinal cord. Neurosurgery 33:277-283; discussion 283-274.
- Krick S, Eul BG, Hanze J, savai R, Grimminger F, Seeger W, Rose F (2005) Role of hypoxia-inducible factor-1alpha in hypoxia-induced apoptosis of primary alveolar epithelial type II cells. Am J Respir Cell Mol Biol 32:395-403.
- Lassmann H (1998) Multiple sclerosis pathology. In: Compston A, editor McAlpine's multiple sclerosis Churchill Livingstone 3rd ed:323-358.
- Lassmann H (2003) Hypoxia-like tissue injury as a component of multiple sclerosis lesions. J Neurol Sci 206:187-191.
- Lauritzen M (1987) Cerebral blood flow in migraine and cortical spreading depression. Acta Neurol Scand Suppl 113:1-40.
- Law M, Saindane AM, Ge Y, Babb JS, Johnson G, Mannon LJ, Herbert J, Grossman RI (2004) Microvascular abnormality in relapsing-remitting multiple sclerosis: perfusion MR imaging findings in normal-appearing white matter. Radiology 231:645-652.

Le Moan N, Baeten KM, Rafalski VA, Ryu JK, Coronado PER,

Bedard C, Syme C, Davalos D, Akassoglou K (2015) Hypocia inducible factor- 1α in astrocytes and/or myeloid cells is not required for the development of autoimmune demyelinating disease. eNeuro 2:1-12.

- Lee SJ, Benveniste EN (1999) Adhesion molecule expression and regulation on cells of the central nervous system. J Neuroimmunol 98:77-88.
- Leng Y, McEvoy CT, Allen IE, Yaffe K (2017) Association of sleep-disordered breathing with cognitive function and risk of cognitive impairment: a systematic review and meta-analysis. JAMA Neurology 74:1237-1245.
- Li L, Welser JV, Dore-Duffy P, Del Zoppo GJ, LaManna JC, Milner R (2010) In the hypoxic central nervous system, endothelial cell proliferation is followed by astrocyte activation, proliferation, and increased expression of the $\alpha 6\beta 4$ integrin and dystroglycan. Glia 58:1157-1167.
- Li L, Welser-Alves JV, van der Flier A, Boroujerdi A, Hynes RO, Milner R (2012) An angiogenic role for the α 5 β 1 integrin in promoting endothelial cell proliferation during cerebral hypoxia. Exp Neurol 237:46-54.
- Lucchinetti C, Brück W, Parisi J, Scheithauer B, Rodriguez M, Lassmann H (1999) A quantitative analysis of oligodendrocytes in multiple sclerosis lesions. A study of 113 cases. Brain 122 (Pt 12):2279-2295.
- Lucchinetti C, Brück W, Parisi J, Scheithauer B, Rodriguez M, Lassmann H (2000) Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. Ann Neurol 47:707-717.
- Lucchinetti CF, Brück W, Rodriguez M, Lassmann H (1996) Distinct patterns of multiple sclerosis pathology indicates heterogeneity on pathogenesis. Brain Pathol 6:259-274.
- Ludwin SK, Johnson ES (1981) Evidence for a "dying-back" gliopathy in demyelinating disease. Ann Neurol 9:301-305.
- Luo F, Liu XJ, Yan N, Li S, Cao G, Cheng Q, Xia Q, Wang H (2006) Hypoxia-inducible transcription factor-1α promotes hypoxia-induced A549 apoptosis via a mechanism that involves the glycolysis pathway. BMC Cancer 6:26.
- Lycke J, Wikkelsö C, Bergh AC, Jacobsson L, Andersen O (1993) Regional cerebral blood flow in multiple sclerosis measured by single photon emission tomography with technetium-99m hexamethylpropyleneamine oxime. Eur Neurol 33:163-167.
- Mahad DH, Trapp BD, Lassmann H (2015) Pathological mechanisms in progressive multiple sclerosis. Lancet Neurol 14:183-193.
- Manoonkitiwongsa PS, Jackson-Friedman C, McMillan PJ, Schultz RL, Lyden PD (2001) Angiogenesis after stroke is correlated with increased numbers of macrophages: The clean-up hypothesis. J Cereb Blood Flow Metab 21:1223-1231.
- Manukhina EB, Goryacheva AV, Barskov IV, Viktorov IV, Guseva AA, Pshennikova MG, Khomenko IP, Mashina SY, Pokidyshev DA, Malyschev IY (2010) Prevention of neurodegnerative damage to the brain in rats in experimental Alzheimer's disease by adaptation to hypoxia. Neurosci Behav Physiol 40:737-743.
- Marrie RA, Reider N, Cohen J, Stuve O, Trojano M, Cutter G, Reingold S, Sorensen PS (2015) A systematic review of the incidence and prevalence of cardiac, cerebrovascular, and peripheral vascular disease in multiple sclerosis. Mult Scler 21:318-331.
- Marshall O, Lu H, Brisset JC, Xu F, Liu P, Herbert J, Grossman RI, Ge Y (2014) Impaired cerebrovascular reactivity in

multiple sclerosis. JAMA Neurol 71:1275-1281.

- McDonald WI (1994) The pathological and clinical dynamics of multiple sclerosis. J Neuropath Exp Neurol 53:338-343.
- McDonald WI, Barnes D (1989) Lessons from magnetic resonance imaging in multiple sclerosis. Trends Neurosci 12:376-379.
- Metz I, Weigand SD, Popescu BF, Frischer JM, Parisi JE, Guo Y, Lassmann H, Brück W, Lucchinetti CF (2014) Pathologic heterogeneity persists in early active multiple sclerosis lesions. Ann Neurol 75:728-738.
- Miller BA, Perez RS, Shah AR, Gonzales ER, Park TS, Gidday JM (2001) Cerebral protection by hypoxic preconditioning in a murine model of focal ischemia-reperfusion. Neuroreport 12:1663-1669.
- Milner R, Crocker SJ, Hung S, Wang X, Frausto RF, Del Zoppo GJ (2007) Fibronectin- and Vitronectin-Induced Microglial Activation and Matrix Metalloproteinase-9 Expression Is Mediated by Integrins α5β1 and αvβ5. J Immunol 178:8158-8167.
- Milner R, Hung S, Erokwu B, Dore-Duffy P, LaManna JC, del Zoppo GJ (2008) Increased expression of fibronectin and the α 5 β 1 integrin in angiogenic cerebral blood vessels of mice subject to hypobaric hypoxia. Mol Cell Neurosci 38:43-52.
- Nathoo N, Rogers JA, Yong VW, Dunn JF (2015) Detecting deoxyhemoglobin in spinal cord vasculature of the experimental autoimmune encephalomyelitis mouse model of multiple sclerosis using susceptibility MRI and hyperoxygenation. PLoS One 10:e0127033.
- Nathoo N, Agrawal S, Wu Y, Haylock-Jacobs S, Yong VW, Foniok T, Barnes S, Obenaus A, Dunn JF (2013) Susceptibility-weighted imaging in the experimental autoimmune encephalomyelitis model of multiple sclerosis indicates elevated deoxyhemoglobin, iron deposition and demyelination. Mult Scler 19:721-731.
- Navarrete C, Carillo-Salinas F, Palomares B, Mecha M, Jimenez-Jimenez C, Mestre L, Feliu A, Bellido ML, Fiebich BL, Appendino G, Calzado MA, Munoz E (2018) Hypoxia mimetic activity of VCE-004.8, a cannabidiol quinone derivative: implications for multiple sclerosis therapy. J Neuroinflammation 15:64.
- Nie XJ, Olsson Y (1996) Endothelin peptides in brain diseases. Rev Neurosci 7:177-186.
- O'Gorman C, Bukhari W, Todd A, Freeman S, Broadley SA (2014) Smoking increases the risk of multiple sclerosis in Queensland, Australia. J Clin Neurosci 21:1730-1733.
- Okuda Y, Sakoda S, Fujimura H, Yanagihara T (2000) The effect of apoptosis inhibitors on experimental autoimmune encephalomyelitis: apoptosis as a regulatory factor. Biochem Biophys Res Comm 267:826-830.
- Ostrow LW, Langan TJ, Sachs F (2000) Stretch-induced endothelin-1 production by astrocytes. J Cardiovasc Pharmacol 36:S274-277.
- Papadaki EZ, Mastorodemos VC, Amanakis EZ, Tsekouras KC, Papadakis AE, Tsavalas ND, Simos PG, Karantanas AH, Plaitakis A, Maris TG (2012) White matter and deep gray matter hemodynamic changes in multiple sclerosis patients with clinically isolated syndrome. Magn Reson Med 68:1932-1942.
- Piechota A, Polańczyk A, Goraca A (2010) Role of endothelin-1 receptor blockers on hemodynamic parameters and oxidative stress. Pharmacol Rep 62:28-34.
- Poyrazoglu HG, Vurdem UE, Arslan A, Uytun S (2016) Evaluation of carotid intima-media thickness in children

with migraine: a marker of subclinical atherosclerosis. Neurol Sci 37:1663-1669.

- Prineas JW, Graham JS (1981) Multiple sclerosis: capping of surface immunoiglobulin G on macrophages engaged in myelin breakdown. Ann Neurol 10:149-158.
- Ranadive SM, Yan H, Weikert M, Lane AD, Linden MA, Baynard T, Motl RW, Fernhall B (2012) Vascular dysfunction and physical activity in multiple sclerosis. Med Sci Sports Exerc 44:238-243.
- Redford EJ, Kapoor R, Smith KJ (1997a) Nitric oxide donors reversibly block axonal conduction: demyelinated axons are especially susceptible. Brain 120 (Pt 12):2149-2157.
- Redford EJ, Smith KJ, Gregson NA, Davies M, Hughes P, Gearing AJ, Miller K, Hughes RA (1997b) A combined inhibitor of matrix metalloproteinase activity and tumour necrosis factor-alpha processing attenuates experimental autoimmune neuritis. Brain 120 (Pt 10):1895-1905.
- Rodriguez M (1992) Central nervous system demyelination and remyelination in multiple sclerosis and viral models of disease. J Neuroimmunol 40:255-263.
- Russ TC, Kivimäki M, Batty GD (2020) Respiratory disease and lower pulmonary function as risk factors for dementia: a systematic review with meta-analysis: Pulmonary function and dementia. Chest epub ahead of print.
- Scannell G, Waxman K, Kaml GJ, Ioli G, Gatanaga T, Yamamoto R, Granger GA (1993) Hypoxia induces a human macrophage cell line to release tumor necrosis factor-alpha and its soluble receptors in vitro. J Surg Res 54:281-285.
- Sixt M, Engelhardt B, Pausch F, Hallmann R, Wendler O, Sorokin L (2001) Endothelial cell laminin isoforms, laminins 8 and 10, play decisive roles in T cell recruitment across the blood-brain barrier in experimental autoimmune encephalomyelitis. J Cell Biol 153:933-945.
- Speciale L, Sarasella M, Ruzzante S, Caputo D, Mancuso R, Calvo MG, Guerini FR, Ferrante P (2000) Endothelin and nitric oxide levels in cerebrospinal fluid of patients with multiple sclerosis. J Neurovirol 6 Suppl 2:S62-66.
- Storch MK, Piddlesden S, Haltia M, Iivanainen M, Morgan P, Lassmann H (1998a) Multiple sclerosis: in situ evidence for antibody- and complement-mediated demyelination. Ann Neurol 43:465-471.
- Storch MK, Stefferl A, Brehm U, Weissert R, Wallström E, Kerschensteiner M, Olsson T, Linington C, Lassmann H (1998b) Autoimmunity to myelin oligodendrocyte glycoprotein in rats mimics the spectrum of multiple sclerosis pathology. Brain Pathol 8:681-694.
- Stowe AM, Altay T, Freie AB, Gidday JM (2011) Repetitive hypoxia extends endogenous neurovascular protection for stroke. Ann Neurol 69:975-985.
- Sun X, Tanaka M, Kondo S, Okamoto K, Hirai S (1998) Clinical significance of reduced cerebral metabolism in multiple sclerosis: a combined PET and MRI study. Ann Nucl Med 12:89-94.
- Swank RL, Roth JG, Woody DC, Jr. (1983) Cerebral blood flow and red cell delivery in normal subjects and in multiple sclerosis. Neurol Res 5:37-59.
- Tabby D, Majeed MH, Youngman B, Wilcox J (2013) Headache in multiple sclerosis: features and implications for disease management. Int J MS Care 15:73-80.
- Tettey P, Simpson S, Jr., Taylor BV, van der Mei IA (2014) Vascular comorbidities in the onset and progression of multiple sclerosis. J Neurol Sci 347:23-33.

Tomasson G, Monach PA, Merkel PA (2009) Thromboembolic

disease in vasculitis. Curr Opin Rheumatol 21:41-46.

- Trapp BD, Stys PK (2009) Virtual hypoxia and chronic necrosis of demyelinated axons in multiple sclerosis. Lancet Neurol 8:280-291.
- Tseng CH, Huang WS, Lin CL, Chang YJ (2015) Increased risk of ischaemic stroke among patients with multiple sclerosis. Eur J Neurol 22:500-506.
- Turnbull IM, Brieg A, Hassler O (1966) Blood supply of cervical spinal cord in man. A microangiographic cadaver study. J Neurosurg 24:951-965.
- Tveten L (1976) Spinal cord vascularity. IV. The spinal cord arteries in the rat. Acta Radiol Diagn (Stockh) 17:385-398.
- Varga AW, Johnson G, Babb JS, Herbert J, Grossman RI, Inglese M (2009) White matter hemodynamic abnormalities precede sub-cortical gray matter changes in multiple sclerosis. J Neurol Sci 282:28-33.
- Wakefield AJ, More LJ, Difford J, McLaughlin JE (1994) Immunohistochemical study of vascular injury in acute multiple sclerosis. J Clin Pathol 47:129-133.
- Wang JS, Liu HC (2009) Systemic hypoxia enhances bactericidal activities of human polymorphonuclear leuocytes. Clin Sci (Lond) 116:805-817.
- Wang Q, Liu ZY, Zhou J (2019) Ultrasonic assessment of carotid intima-media thickness in migraine: a metaanalysis. J Int Med Res 47:2848-2855.
- Weinstock-Guttman B, Zivadinov R, Mahfooz N, Carl E, Drake A, Schneider J, Teter B, Hussein S, Mehta B, Weiskopf M, Durfee J, Bergsland N, Ramanathan M (2011) Serum lipid profiles are associated with disability and MRI outcomes in multiple sclerosis. J Neuroinflammation 8:127.
- Weinstock-Guttman B, Zivadinov R, Horakova D, Havrdova E, Qu J, Shyh G, Lakota E, O'Connor K, Badgett D, Tamaño-Blanco M, Tyblova M, Hussein S, Bergsland N, Willis L, Krasensky J, Vaneckova M, Seidl Z, Ramanathan M (2013) Lipid profiles are associated with lesion formation over 24 months in interferon-β treated patients following the first demyelinating event. J Neurol Neurosurg Psychiatry 84:1186-1191.
- Wingerchuk DM, Carter JL (2014) Multiple sclerosis: current and emerging disease-modifying therapies and treatment strategies. Mayo Clin Proc 89:225-240.
- Yaffe K, Laffan AM, Harrison SL, Redline S, Spira AP, Ensrud KE, Ancoli-Israel S, Stone KL (2011) Sleep-disordered breathing, hypoxia, and risk of mild cognitive impairment and dementia in older women. JAMA 306:613-619.
- Yang R, Dunn JF (2015) Reduced cortical microvascular oxygenation in multiple sclerosis: a blinded, casecontrolled study using a novel quantitative near-infrared spectroscopy method. Sci Rep 5:16477.
- Yohannes AM, Chen W, Moga AM, Leroi I, Connolly MJ (2017) Cognitive Impairment in Chronic Obstructive Pulmonary Disease and Chronic Heart Failure: A Systematic Review and Meta-analysis of Observational Studies. J Am Med Dir Assoc 18:451.
- Yuksel B, Koc P, Ozaydin Goksu E, Karacay E, Kurtulus F, Cekin Y, Bicer Gomceli Y (2019) Is multiple sclerosis a risk factor for atherosclerosis? J Neuroradiol.
- Zeis T, Probst A, Steck AJ, Stadelmann C, Brück W, Schaeren-Wiemers N (2009) Molecular changes in white matter adjacent to an active demyelinating lesion in early multiple sclerosis. Brain Pathol 19:459-466.
- Zhen J, Wang W, Zhou J, Qu Z, Fang H, Zhao R, Lu Y, Wang H, Zang H (2014) Chronic intermittent hypoxic

preconditioning suppresses pilocarpine-induced seizures and associated hippocampal neurodegeneration. Brain Res 1563:122-130.